Systematic Reviews and Harm-Benefit Assessment, Voss 27-28 May 2015

Where can we find 3R literature?

Adrian Smith adrian.smith@norecopa.no



www.norecopa.no

- What is 3R literature?
- Why is it hard to find?
- What can we do about?
- Examples of 3R sources
- Tools for searching the literature



Reporting

Planning







3R literature embraces all 3R alternatives

1) Replacement alternatives

3D models Audiovisual aids **Computer simulations** Mannekins, simulators, robots QSAR (Quantitative Analysis of Structure/Activity Relationships) Cell and tissue cultures, organoids, organ perfusion High Throughput Screening (HTS), organs-on-a-chip Biochemical & immunological methods (RIA, ELISA) Hybrid DNA technique, GMM Trials on "lower" organisms Acute experiments (terminal anaesthesia) Trials on dead animals Trials on humans (microdosing and medical imaging) Synthesis of new evidence from experiments that have already been performed



High Throughput Screening (HTS)





Robotic platform with high-throughput liquid handler for sample dilution and treatment. Optical plate reader and incubator



Cell culture and compound management

Data management system

Automated imaging microscope for highcontent screening

https://eurl-ecvam.jrc.ec.europa.eu/laboratories-research/high-throughput-screening-and-test-development/hts



Lung-on-a-chip Wyss Institute, Harvard



Next generation Multi-Organ-Chip





Marx et al., Altern Lab Anim. 2012 Oct;40(5):235-57



- Norecopa -

Integrating natural science and technology: Fish and fish robots







photo: Norecopa

Prof. Maarja Kruusma







Photo: Joe Mcdonald/ Corbis

http://www.theguardian.com/commentisfree/2012/mar/14/laboratories-animals-anti-vivisection-campaign

Fidelity: overall proportionate difference (e.g. HiFi)

Discrimination: the extent to which the model reproduces one particular property in which we are interested



www.frame.org.uk/tag/russell-and-burch





http://www.interniche.org/ko/node/5134

https://www.wardsci.com/store/catalog/product.jsp? catalog_number=813015#

The potential for 3R alternatives cannot be evaluated until the objective of the study is known. This applies to all use of animals in research, testing, education and training

Possible objectives in education & training:

- Teaching and practising:
 - laboratory skills
 - general animal handling skills
 - preparation-specific animal skills
- imparting good ethical thinking
- new knowledge and reinforcing existing
- data handling skills
- experimental design skills
- communication skills (oral, written)
- groupwork
- staff-student interaction

AJ Smith & K Smith, 2004

Guidelines for humane education: Alternatives to the use of animals in teaching and training

Proceedings of the 4th World Congress on Alternatives and Animal Use in the Life Sciences, New Orleans, August 2002.

http://www.atla.org.uk/wp-content/plugins/s2member-files/ 32_S1a_3_Plenary_specialcontribution.pdf (log-in required, pages 16-26)

Where do I find information about alternatives for use in Education and Training?

Databases NORINA (oslovet.norecopa.no/NORINA) InterNICHE (interniche.org/en/alternatives)

Loan system InterNICHE (interniche.org/en/loansystem)

Do we need an alternative?



http://www.all-creatures.org/anex/cat-res-07.html

3) Reduction alternatives

A good statistician is the lab animal's best friend.

Combined with methods to reduce background "noise".



http://norecopa.no/norecopa/vedlegg/Berdoy-handout.pdf

Sources of background "noise":

- Age, sex, weight
- Stress, subclinical disease
- Room temperature, animal cage
- Environmental "enrichment"
- Temporal differences between treatments
- Climatic factors
- Position of cage in the room
- Experimenter
- Animal Technician (weekend workers)
- and many more

3) Refinement alternatives

"Simple" techniques?



Photo: NMBU

Are they feasible? e.g. i.m. injections

"Simple" identification methods? Do they affect the animal?



Photo: T. Poppe, NMBU



http://blogs.discovermagazine.com/notrocketscience/2011/01/12/ flipper-bands-impair-penguin-survival-and-breeding-success/ #.VLU6_8Y7_wo



Photo: colourbox.com

Refinement to avoid **contingent suffering**

(not just direct suffering caused by the procedure)

e.g. fear, boredom, discomfort

which may caused by

e.g. transport, housing, husbandry, social hierarchy

Single-housed male mice show symptoms of what in humans would be characterised as depression

http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0111065



An useful additional (but largely unknown) tool... Carol M. Newton (1925-2014)



National Library of Medicine

The three S's

- Good Science
- Good Sense*
- Good Sensibilities*

*We can do this ourselves without scientific literature!

Carol M Newton, quoted in Rowsell HC (1977): The Ethics of Biomedical Experimentation in The Future of Animals, Cells, Models, and Systems in Research, Development, Education, and Testing pp. 267-281, National Academy of Sciences, Washington, D.C., ISBN 0-309-02603-2.



f The 3Rs

Our science

Funding

3Rs resources

nding

Home > News > Blog > Creating a culture of care

Creating a culture of care

Friday 22 August 2014

Dr Marilyn Brown, Corporate Vice President of Global Animal Welfare at the contract research organisation Charles River, has many years of experience managing experimental facilities and animal care programmes.

https://www.nc3rs.org.uk/news/creating-culture-care



Establishing a Culture of Care, Conscience, and Responsibility: Addressing the Improvement of Scientific Discovery and Animal Welfare Through Science-based Performance Standards

H. J. Klein and K. A. Bayne

Address correspondence and reprint requests to Dr. Klein, Merck Research Laboratories, WP42-211, West Point, PA 19486, or email Hilton_klein@merck.com.

http://ilarjournal.oxfordjournals.org/content/48/1/3.full





Reporting

Planning







Why is 3R literature hard to find?

- Bibliographic databases are often not used adequately (poor overlapping between the databases)
- Too few scientists are aware of the specialist 3Rdatabases
- Scientists rarely use "3R" words when they write titles/abstracts/keywords for their papers
- Databases rarely flag 3R-papers with explicit thesaurus terms ^(B)
- We have no single "Journal of Alternatives"

Reporting has historically been poor:

Jane Smith et al. (1997): 149 papers in 8 journals from 1990-1991:

Parameters <u>not</u> mentioned:

Number of animals: 30%

How the animals were killed: 45%

Sex	28%	Room temperature	72%
Age	52%	Relative humidity	89%
Weight	71%	Photoperiod	72%
Source	53%	Number of animals/cage	73%

Often detailed descriptions of chemicals, equipment and treatments, but very little about the animals, choice of sample size, randomisation etc:

'white mice were used'

Many of these omissions make it harder to advance the 3Rs, e.g.

- *methods of drug administration and blood sampling*
- details of anaesthesia and analgesia
- humane endpoints

Kilkenny C et al. (2009)

271 papers, mostly in 2003-2005

http://www.plosone.org/article/info%3Adoi%2F10.1371%2Fjournal.pone.0007824

Many studies did not

- *describe the animals adequately*
- describe how the sample size was chosen
- describe how the animals were allocated to the treatment groups, and whether the observations were performed blind.

Even the titles, keywords and abstracts are often not very informative and lack 3R terms

The development of Response Surface Pathway Design in toxicity studies

The development of Response Surface Pathway Design to reduce animal numbers in toxicity studies

3R methods are often not highlighted in the scientific literature



http://www.theodora.com/rodent_laboratory/ blood_collection.html



photo:NMBU

SCID-Hu mice immunized with a pneumococcal vaccine produce specific human antibodies and show increased resistance to infection. Saphenous vein puncture for blood sampling of the mouse, rat, hamster, gerbil, guinea-pig, ferret and mink

Visibility! Not necessarily in a high-impact journal.



Part of the problem:

Reporting (the Materials & Methods section) should ideally be so detailed that it is possible to reproduce the study in another lab.

But this information takes space.

Although space is limited, we waste space...

'drinking' water

'farm' pigs

'under approved conditions' (who approved them?) 'housed under standard conditions' 'given analgesia' Many journals now offer supplementary online space (generally unlimited) where more information about the methods and results can be posted.

And most people have access to a website where this could be posted...

Publication of negative results

- Bias automatically occurs if only positive results are reported!
- Negative results may be just as important for the scientific community, even if they are less newsworthy
- Many medical journals require registration of trials before they start, to prevent the under-reporting of negative results (http:// www.icmje.org/recommendations/browse/publishing-and-editorialissues/clinical-trial-registration.html)

There are a number of journals now that report negative results, e.g.

J of Negative Results (http://www.jnr-eeb.org/index.php/jnr) J of Negative Resuts in Biomedicine (http://www.jnrbm.com) J of Pharmaceutical Negative Results (http://www.pnrjournal.com) The All Results Journals (http://www.arjournals.com/ojs)



Reporting Planning Research

Identify and ensure the quality of (at least) the critical points in the experiment: critical for animal welfare and scientific value

Good reporting and increased implementation of the 3Rs is dependent upon:

Quality assurance and a culture of care at all levels of the animal facility.

- SOPs describing good techniques, carried out by competent operators
- A Checklist ("contract") between researcher and the facility
- The AAALAC Program Description template or something similar as an overall quality assurance checklist for the facility
- A Master Plan as a weekly checklist for the whole facility

Template for a Program Description from AAALAC International

- Animal care and use policies and responsibilities
- Animal environment, housing and management
- Veterinary medical care
- Physical plant

https://www.aaalac.org/programdesc/index.cfm



AAALAC. Where science and responsible animal care connect.



www.aaalac.org

OUTLINE

DESCRIPTION OF INSTITUTIONAL ANIMAL CARE AND USE PROGRAM

- I. Introduction
 - A. Name of Program Unit
 - B. Overview and Purpose
 - C. <u>Description of the Organization</u> (Attach organizational chart plus any support comments needed)
 - D. Key Institutional Representatives
 - E. Accreditation History
 - F. Nature of Research, Testing, and Teaching Programs
 - G. Research Funding Source(s)
 - H. Summary of Facilities
 - I. Other Units not Included in This Description
 - J. Contract Facilities
 - K. Other Relevant Background
- II. Description
 - A. Institutional Policies and Responsibilities
 - 1. Monitoring the Care and Use of Animals
 - a. Institutional Animal Care and Use Committee (s) (IACUC)
 - 1) Who appoints Committee/who is Institutional Official
 - 2) Composition/Frequency of Meetings/Responsibilities of the Committee

10/97



Guidelines

e.g.

- ARRIVE, ILAR, ICLAS, LASA/APC
- GSPC
- · Guidelines for specific types of animal research

1000	⊋ Enable synonyms and stemming Reset
Database:	Search in:
3R Guide Classic AVs. NORINA	All Text Title Author Publisher Supplier
TextBase	Record Number

Guidelines for reporting the results of experiments on fish 3R Guide/15075

A detailed account of experimental design, including an accurate description of the animals used, is an essential part of good research practice. This paper suggests guidelines for reporting fish experiments.

Guidelines for reporting the results of experiments using mammals 3R Guide/15076

A detailed account of experimental design, including an accurate description of the animals used, is an essential part of good research practice. This paper suggests guidelines for reporting experiments on mammals.

Recommendations for the conduct, reporting, editing and publication of scholarly work in medical journals

3R Guide/15093

The International Committee of Medical Journal Editors (ICMJE) developed these

Category

Agricultural animals (8)

- Anaesthesia and analgesia (12)
- Aquatic animals (5)
- Behavioural research (6)
- Birds (7)
- Blood sampling (8)
- Cancer research (5)
- Design (31)
- Disease research (7)
- Education and training (10)
- Environmental enrichment (9)
- T Ethics & harm-benefit analysis (17)
- Fish (8)
- Handling (14)
- Housing and management (27)
- Humane killing (15)
- Miscellaneous (1)
- ¬ Neuroscience research (2)
- n Non-human primates (12)
- n Nutritional research (1)
- Procedures (30)
- Reporting (19)
- Surgical research (10)
- Toxicology (6)
- Transport (8)
- Wildlife (8)

Interview Provide as accurate simulation concers a description of the content of the another sposes bits Instruct Provide an accurate summary of the background, research objectives, including details of the species or strain of animal used, key methods, principal findings and concessions of the strain species or strain of animal used, key methods, principal findings and concessions of the species of strain of experimental approach and rationale. Provide an work in the experimental approach and rationale. b. Experimental approach and rationale. D. Experimental approach and rationale. b. Experimental approach and rationale. D. Experimental approach and rationale. b. Experimental approach and rationale. Introducts the nature of the sthreal new permissions, relevant locances in g Arman (Scientrike Procedures) Act 1986, and national or institutional guidelines for the care and use of nimes, that cover the research. Indicate the nature of the sthreal new interfaces of subjective bias, when assessing results a indicate the nature of the strain institutional groups. b. Any steps taken to minimize the strate of subjective bias, when assessing results a indicate scrifted built. Indicate the nature of the spectromental indicate or insubjective bias, when assessing results a indicate scrifted built. b. Any steps taken to minimize the strate of animabile. Indicate the nature of the scrifted built on thustater now complex study designs were carried built. b. Any steps taken to minimize the strate of animabile. Indicate the nature of the strate on the	11 1.00	
Instruct Provide an accurate summary of the background, research objectives, including details of the species or strain of an initial used, key including, principal findings and boncurates and the study. VICUUCTION a. Include sufficient scientific background [including rolevant references to previous work to understation and contexts for the study, and explain the experimental approach and rationale. b. Explain now and why the animal used is an address the scientific to the study and explain the experimental approach and rationale. b. Explain now and why the animal species and model being used can address the scientific hypotheses being tested. Interaction Clearly cascriber the primary and any secondary objectives of the study or seecific hypotheses being tested. Interaction Indicate the nature of the ethical review permissions, reviewant licences (e.g. Anima [Scientific Procedures] Act 1986], and national or institutional guidelines for the case and use of animals. It that cover the research is reviewed and when assessing results in the scientific Procedures (a the study design including:	line	as possible.
An exploremental and the experimental scient file background linchading reavant references to previous write to understand the embination and contact for the study, and explain the experimental approach and rationale. Belground a. Include sufficient scient file background linchading reavant references to previous write to understand the embination and contact for the study, and explain the experimental approach and rationale. Belground a. Explain now and why the animal species and model being used can address the actient file dogethwas and where appropriate, the study is relevance to than all bology. Anternal Scientific Procedures Act 1986, and national or institutional guidelines for the care and use of animals, that cover the research. Indicate the nature of the ethical review permissions. relevant licences (e.g. Annual (Scientific Procedures) Act 1986), and national or institutional guidelines for the care and use of animals, that cover the research. Itudy design a. The number of experimental and control groups. b. Any steps taken to minimise the efflects of subjective bas, when elecating animal sto to earlie of animals. A time-line diagram of low chart can be useful to illustrate now complex study design results is all folders. c. The experimental and leca a single animal, group or cage of animals). A time-line diagram of low chart can be useful to illustrate now complex study design including: a study design for uding controls, provide procedures and group including surgical processing, including surgical processing, including surgical processing, and stops desploy of animals). A time-line diagram	bstract	Provide an accurate summary of the background, research objectives, including details of the species or strain of animal used, key methods, principal findings and conclusions of the study.
addrouble 3 a. Include sufficient scientific background [including mawart references to previous work to understand the motivation and context for the study, and explain the experimental approach and rationals. b. Explain now and why the animal species and model being used can address the scientific objectives and, where appropriate, the study's relevance to humanic background b. Explain now and why the animal species and model being used can address the scientific objectives and, where appropriate, the study is relevance to humanic background b. Explain now and why the animal species and model being used can address the subdy? b. Explain now and why the animal species and model being used can address the subdy? b. Explain now and why the animal species and model being used can address the subdy? b. Explain now and why the animal species and model being used can address the subdy? b. Explain now and why the animal species and model being used can address the subdy? b. Explain now and why the animal species and model being used can address the subdy? b. Explain now and why the animal species and model being used can address the subdy? b. Explain now and why the animal species and model being used can address the subdy? b. Indicate the nature of the ethical review permissions. relevant licences (e.g. Amma (Scientife Procedures Art 1966), and national subdy design including: b. Any steps taken to minimiss the effects of subjective bias, when allocating animals to treatment (g.g. address thespecimenta), and when asserting results in g.g. from explainteri.	TRODUCTION	
b. Explain how and why the animal species and model being used can address the scient file objectives and, where appropriate, the study's relevance to human biology. b. Explain how and why the animal species and model being used can address the scient file objectives and, where appropriate, the study's relevance to specific hypotheses being tested. b. Explain how and why the animal species and model being used can address the specific hypotheses being tested. b. Explain how and why the animal species and model being used can address the specific hypotheses being tested. b. Explain how and why the animal species and model being used. b. Explain how and why the animal species and model being used. b. Explain how and why the animal species and model being used can address the specific hypotheses being tested. b. Explain how and why the animal species and model being used (an address the specific hypotheses). b. Explain how and why the animal species and model being used. b. Explain how and why the animal species and model being used. b. Explain how and why the animal species and model being used. b. Explain how and why the animal species provide provide animals. b. Explain how and why the animal species and model being used. b. Explain how and why the animals the effects of subjective bas when allocating animals is the species of animals. b. Any steps taken to minimize the experimental group, including controls, provide procedures the care and use of all procedures to reade of animals. <	ackground 3	a. Include sufficient scientific background (including relevant references to previous work) to understand the motivation and context for the study, and explain the experimental approach and rationale.
A Clearly describe the primary and any secondary objectives of the study or specific hypotheses being tested. IFTHODS thick al statement 5 Indicate the nature of the ethical review permissions, relevant licences (e.g. Anima) (Scientric Procedures) Acti 1986), and national or institutional guidelines for the care and use of animals, that cover the research. tudy design 6 For each experiment, give brief details of the study design including: Any steps taken to minimize the effects of wubjective bias, when elecating animals to treatment (e.g., rendomisation procedure) and when assessing results is g. if done, describe who was blinded and when. The experimental unit (e.g. a single animal, group or cage of animals). A time-line diagram of flow chiert can be useful to illustrate how complex study designs were carried out. Revelues 7 For each experiment and each experimental group, including controls, provide procedures tarried out. Revelues 7 For each any formulation and close, size and route of administration, aniesthesia and analgosia used (including monitoring) surgical procedure, method of orderansals. Provide details of any specialist equipment used, including surgical procedure, method of orderansals. Provide details of any specialist equipment used, including surgical procedure, animational gene used). When (e.g. time of day). When (e.g. time of day). When (e.g. time of day). When (e.g. time of choice of specific ansee		b. Explain how and why the animal species and model being used can address the acientific objectives and, where appropriate, the study's relevance to human biology.
EFFHOOS Indicate the nature of the ethical review permissions, relevant licences (e.g. Animal [Scientific Procedures] Act 1986], and national or institutional guidelines: for the care and use of animals, that cover the research. tudy design 6 For each experiment, give brief details of the study design including: a. The number of experimental and control groups. b. Any steps taken to minimise the effects of subjective bias when electaing animals to breatment (e.g., randomisation procedure) and when assessing results is g. if done, describe who was blinded and when). c. The experimental until leg a single animal, group or cage of animata). A time-line diagram of flow chart can be useful to illustrate now complex study designs were carried out. Reperimental cocedures 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. End was blinded and when? 2 For each graphic animals. 3 A time-line diagram of flow chart can be useful to illustrate now complex study designs were carried out. 5 End of euthanssial. Provide details of any specialist equipment used. Including supplicity. 6 When (e.g. time of day). C. Where (e.g. time of day). 6. Where (e.g. time of day). 6. Where (e.g. time of day). 7. Where (e.g. time of day). C. Where (e.g. time of the animals used including species strain, sex. developmental steps (e.g. mean or median ege plus age	Nectives 4	Clearly describe the primary and any secondary objectives of the study, or specific hypotheses being tested.
Indicate the nature of the ethical review permissions, relevant licences (e.g. Animal [Scientific Procedures] Act 1986], and national or institutional guidelines for the care and use of animals, that cover the research. tudy design 6 For each experiment, give brief details of the study design including: a. The number of experimental and control groups. b. Any steps taken to minimise the effects of subjective bias when elecating animals to breatment (e.g. randomisation proceedure) and when assessing results is g. if done, describe who was blinded and when?. c. The experimental unit (e.g. a single animal, group or cage of animals). A time-line diagram or flow chart can be useful to illustrate now complex study designs were carried out. Reperimental coeffures 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. Error example: a. How (e.g. time formulation and dose, site and route of administration, aniesthesia and analgesia used [including monitoring], surgical procedure, method of outhanasial. Provide details of any specialist equipment used, including supplier(s). E. When (e.g. time of day! E. When (e.g. time of day! e. When (e.g. time of day! Provide distails of the anima's used, including species, strain, sex, method of subanasial. Provide details of any specialist equipment used, including supplier(s). E. When (e.g. time of day! E. When (e.g. time of day! Provide distails of the anima's used, including species, strain, sex, method or subanas (e.g. more rouging spe	ETHODS	
tudy design 6 For each experiment, give brief details of the study design including: a. The number of experimental and control groups. b. Any steps taken to minimise the effects of subjective bias when allocating animals to breatment (e.g. randomisation procedure) and when assessing results lag. If done, describe who was blinded and when). c. The experimental unit (e.g. a single animal, group or cage of animals). A time-line diagram or flow chart can be useful to illustrate how complex study designs were carried out. For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. For example: a. How (e.g. drug formulation and dose, site and route of administration, aniesthesis and analgesia used (including monitoring), surgical procedure, method of euthansial. Provide details of any specialist equipment used, including supplicits! b. When (e.g. time of day). c. Where (e.g. home cage, altoratory, water mazo). d. Why (e.g. rotionals for choice of specific ansesthetic, route of administration, drug dose used). Experimental and provide details of the anima is used, including species, strain, sex, device details of the anima is used, including species, strain, sex, device, mean or median weight range). b. Provide further relevant information such as the source of administration, leg, mean or median weight range). b. Provide further relevant information such as the source of animals, limitation isor moment and experimental useful range). b. Provide further relevant information such as the source of animals, limitation momention such as the source of animals.	hical statement 5	Indicate the nature of the ethical review permissions, relevant licences (e.g. Animal [Scientific Procedures] Act 1986), and national or institutional guidelines for the care and use of animals, that cover the research.
a. The number of experimental and control groups. b. Any steps taken to minimise the effects of subjective bass when allocating animals to breatment (e.g., randomisation procedure) and when assessing results is g. if done, describe who was blinded and when. c. The experimental unit leng, a single animal, group or cage of animals). A time-line diagram or flow chart can be useful to illustrate now complex study designs were carried out. experimental rocedures 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. For example: a. How (e.g. drug formulation and dose, size and route of administration, anaresthasia and analgesia used (including monitoring), surgical procedure, method of eithanasial. Provide details of any specialist equipment used. Including supplicits! b. When (e.g. time of day)! c. Where (e.g. home cage, laboratory, water maze). d. Why (e.g. notice of specific anaesthetic, route of administration, drug dose used). a. Provide details of the anima is used. including species, strain, sex, anivalization, drug dose used). xperimental 8 a. Provide fact anima is used. including species, strain, sex, anivalization, and species when are precise, and weight range). b. Provide fact weight prus weight range). b. Provide fact is of the anima is used. including species, strain, sex, anivalization, drug dose used). experimental a. Provide fact and weight prus weight range). b. Provide further re	tudy design 6	For each experiment, give brief details of the study design including:
b. Any steps taken to minimise the effects of subjective bias when allocating animals to treatment (e.g. randomisation procedure) and when assessing results it a, if done, describe who was blinded and when). c. The experimental unit (e.g. a single animal, group or clige of animats). c. The experimental unit (e.g. a single animal, group or clige of animats). A time-line diagram or flow chart can be useful to illustrate now complex study designs were carried out. xperimental increases 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. Kperimental increases 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. Kperimental increases 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. Kperimental increases 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. Kperimental increases 7 For each experiment and each experimental group, including controls, provide including supplicits. b. When (e.g. thing of formulation and dose, site and route of administration, anitoting, surgical procedure, method of euthanasial. Provide details of any specialist equipment used. Including subjective, water mazel. c. Where (e.g. home cage, laboratory, water mazel. d. Why (e.g. rotionale for choice of specific ansesthetic, route of administration, drug dose used		a. The number of experimental and control groups.
c. The experimental unit log, a single animal, group or cage of animals). A time-line diagram or flow chart can be useful to Illustrate how complex study designs were carried out. experimental noted and experiment and each experimental group, including controls, provide precise details of all procedures carried out. For example: a. How (e.g. drug formulation and dose, site and route of administration, anaesthesis and analgesia used (including monitoring), surgical procedure, method of exthansial, Provide details of any specialist equipment used, including supplicifs). b. When (e.g. time of day). c. Where (e.g. home cage, laboratory, water maze). d. Why (e.g. rotionals for choice of specific anaesthetic, route of administration, drug dose used). experimental 8 a. Provide datals of the animals used, including species, strain, sex, anivaling mean or median age plus age range) and weight lieg, mean or median weight range). b. Provide further relevant information such as the source of administration relevance in the animals used in cuding species, strain, sex, devisionment is strain moment atus, genetic modification status (e.g. wrock-out or test precision in the animals used including species, strain, sex, devisionment is a strain moment atum, genetic modification status (e.g. wrock-out or test precision in the animals used in formation such as the source of administration or test precision in the animals weight range).		b. Any steps taken to minimise the effects of subjective bias when allocating animals to treatment (e.g. randomisation procedure) and when assessing results (e.g. If done, describe who was blinded and when).
A time-line diagram of flow chart can be useful to Illustrate now complex study designs were carried out. xperimental rocedures 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. For example: a. How (e.g. drug formulation and dosa, site and route of administration, anaesthesia and analgesia used (including monitoring), surgical procedure, method of euthanasial, Provide details of any specialist equipment used. Including supplier(s). b. When (e.g. time of day). c. Where (e.g. home cage, laboratory, water maze). d. Why (e.g. rotionale for choice of specific anaesthetic, route of administration, drug dose used). xperimental instals 8 a. Provide details of the animals used, including species, strain, sex, development a stuge (e.g. mean or median age plus age range) and weight le.g. mean or median weight range). b. Provide further relevant information such as the source of administration previde method information such as the source of animals, international strain momento stude, genetic modification status (e.g. wrock-out presensence)		c. The experimental unit (e.g. a single animal, group or cage of animals)
xperimental rocedures 7 For each experiment and each experimental group, including controls, provide precise details of all procedures carried out. For example: a. How (e.g. drug formulation and dose, site and route of administration, anaristhesia and analgesia used (including monitoring), surgical procedure, method of euthanasial, Provide details of any specialist equipment used. Including supplier(s). b. When (e.g. time of day). c. Where (e.g. home cage, laboratory, water maze). d. Why (e.g. rotionale for choice of specific anaesthetic, route of administration, drug dose used). xperimental autistal 8 a. Provide details of the animals used, including species, strain, sex, developmental stage (e.g. mean or median age plus age range) and weight le.g. mean or median weight range). b. Provide further relevant information such as the source of administration procedure is train momental true, genetic modification status (e.g. wrock-out or transport in accestion).		A time-line diagram or flow chart can be useful to illustrate how complex study designs were carried out.
For example: a. How (e.g. drug formulation and dose, site and route of administration, anaesthesis and analgesia used (including monitoring), surgical procedure, method of exthansial, Provide details of any specialist equipment used, including supplicr(s). b. When (e.g. time of day). c. Where (e.g. time of day). c. Where (e.g. time of day). d. Why (e.g. rotionale for choice of specific anaesthetic, route of administration, drug dose used). xperimental 8 a. Provide details of the animals used, including species, strain, sex, newslopment a stage (e.g. mean or median age plus age range) and weight leag, mean or median weight rangel. b. Provide further relevant information such as the source of administration relevance in general strain moment at use of the animals weight rangel.	xperimental 7 rocedures	For each experiment and each experimental group, including controls, provide precise details of all procedures carried out.
b. When (e.g. time of day). c. Where (e.g. home cage, laboratory, water maze). d. Why (e.g. rationale for choice of specific anaesthetic, route of administration, drug dose used). xperimental xperimental 8 a. Provide details of the animals used, including species, strain, sex, developmental stage (e.g. mean or median age plus age range) and weight lieg, mean or median weight range). b. Provide further relevant information such as the source of animals, international strain nomencature, genetic modification status (e.g. whock-out or beforem the animal strain nomencature, genetic modification status (e.g. whock-out or beforem the animal strain or median weight range).		For example: a. How (e.g. drug formulation and dose, site and routs of administration, anaesthesis and analgesis used (including monitoring), surgical procedure, method of euthanasia). Provide details of any specialist equipment used, including supplier(s).
c. Where (e.g. home cage, laboratory, water maze). d. Why (e.g. rationale for choice of specific anaesthetic, route of administration, drug dose used). xperimental 8 a. Provide details of the animals used, including species, strain, sex, developmental stage (e.g. mean or median age plus age range) and weight la.g. mean or median weight plus weight range). b. Provide further relevant information such as the source of animals, immational strain nomenciature, genetic modification status (e.g. knock-out or transported nearborn status (e.g. knock-ou		b. When (e.g. time of day).
d. Why is g. rationals for choice of specific anaesthetic, route of administration, drug dose used).		c. Where (e.g. home cage, laboratory, water maze).
xperimental 8 a. Provide details of the animals used, including species, strain, sex, developmental stage (s.g. mean or median age plus age range) and weight likig, mean or median weight plus weight range). b. Provide further relevant information such as the source of animals, international strain nomencature, genetic modification status (e.g. knock-out or bespecies).		d. Why is g. rationals for choice of specific anaesthetic, route of administration, drug dose used).
b. Provide further relevant information such as the source of animals. International strain nomenciature, genetic modification status (e.g. knock-out or transported account and the animatic trains of the or test pairs and previous of the status of the status of the status of the or test pairs of the original status of the status of the original status of the ori	xperimental 8 niztala	a. Provide details of the animals used, including species, strain, sex, developmental stage (e.g. mean or median age plus age range) and weight (e.g. mean or median weight plus weight range).
procedures, etc.		b. Provide further relevant information such as the source of animals, imemational strain nomenciature, genetic modification status (e.g. knock-out or transgenic), genotype, health/immune status, drug or test naive, previous procedures, etc.

illidellr	<u>1es</u>	
	.00	ide details of:
in the second		a. Housing itype of facility e.g. specific pathogen free (SPF); type of cage or housing; bedding material; number of cage companions; tank shape and material atc. for fash.
		b. Husbandry conditions (e.g. breeding programme, light/dark cycle, temperature, quality of water etc for fish, type of food, access to food and water, environmental enrichment).
		c. Welfare-related assessments and interventions that were carried out prior to, during, or after the experiment.
Sample size	10	a. Specify the total number of animals used in each experiment, and the number of animals in each experimental group.
		b. Explain how the number of animals was arrived at. Provide details of any sample size calculation used.
		c. Indicate the number of independent replications of each experiment, if relevant
Allocating animals o experimental	11	 Give full details of how animals were allocated to experimental groups, including randomisation or matching if done.
roupa		b. Describe the order in which the animals in the different experimental groups were treated and assessed.
aperimental atcomes	12	Clearly define the primary and secondary experimental outcomes assessed (e.g. cell death, molecular markera, behavioural changes):
Statistical methods	13	a. Provide details of the statistical methods used for each analysis.
		b. Specify the unit of analysis for each dataset (e.g. single animal, group of animals, single neuron).
		c. Describe any methods used to assess whether the data mit the assumptions of the statistical approach.
RESULTS		
iaaeline data	14	For each experimental group, report relevant characteristics and health status of animals (e.g. weight, microbiological status, and drug or test naive) prior to treatment or testing (this information can often be tabulated).
lumbers analysed	15	 Report the number of animals in each group included in each analysis. Report absolute numbers (e.g. 10/20, not 50%²).
		b. If any animals or data were not included in the enalysis, explain why.
Jutcomes and stimation	16	Report the results for each analysis carried out, with a measure of precision (e.g. standard error or confidence interval).
Adverse eventa	17	a. Give details of all important adverse svents in each experimental group.
		b. Describe any modifications to the experimental protocols made to reduce adverse events.
DISCUSSION		and the second se
nterpretation/	18	a. Interpret the results, taking into account the study objectives and hypotheses, current theory and other relevant studies in the literature.
		.b. Comment on the study ilmitations including any potential sources of bias, any limitations of the animal model, and the imprecision associated with the results ² .
		c. Describe any implications of your experimental methods or findings for the replacement, refinement or reduction (the 3Rs) of the use of animals in research.
Teneralisability/	19	Comment on whether, and how, the findings of this study are likely to translate to other species or systems, including any relevance to human biology.
Punding	20	List all funding sources (including grant number) and the role of the funder(s) in the study.

https://www.nc3rs.org.uk/arrive-guidelines

The ARRIVE guidelines

Carol Kilkenny¹, William J Browne², Innes C Cuthill³, Michael Emerson⁴ and Douglas G Altman⁵

¹The National Centre for the Replacement, Refinement and Reduction of Animals in Research, London, UK, ²School of Veterinary Science, University of Bristol, Bristol, UK, ³School of Biological Sciences, University of Bristol, Bristol, UK, ⁴National Heart and Lung Institute, Imperial College London, UK, ⁵Centre for Statistics in Medicine, University of Oxford, Oxford, UK

Animal Research: Reporting of In Vivo Experiments

ARRIVE (动物研究: 体内实验报告) 指南是由国家3R中心创设,旨在通过提高动物研究设计,分析和报告的质量,使 报告的信息量最大化并将不必要的研究减至最低程度。 该指南于2010年6月在PLOS Biology 网络杂志发表,并得 到多家科研杂志、主要的资助机构和学术团体的赞同。

ARRIVE 指南的宗旨是:	ARRIVE 指南无意于:	ARRIVE指南适用于哪些科研领域?	如何使用ARRIVE 指南?	· · · · · · · · · · · · · · · · · · ·
 提高动物研究报告的质量。 指导作者在稿件中提供必要的信息,但并不是硬性规定。 报告的灵活性使之广泛适应于各种研究领域和实验方案 促进具有可重复性、透明性、精确性、全面性,、简明性、逻辑性的高质量论文。 促进科研成果在科学界更广泛的交流。 	 促进统一性, 扼杀创造性, 或鼓励 作者条条框框。 有些条款并不适 用于所有的研究, 有些可用图表及 说明或流程图展示 (如所处理的, 评估的和分析的动物数量)。 提供实验设计和执行的指南。 但 是指南中有些条款譬如随机化, 施 盲和使用对照组等对于设计实验 时减少偏倚风险和提高研究的稳 健性是有帮助的 	 最适用于比较研究,即两个或多个 实验动物组进行比较,其中一组或 多组常设为对照组。也适用于比 较药物不同剂量的研究,或者如用 单一动物作为其自身对照 (被试内 实验)。 大多数建议也适用于不含对照组的 研究。 适用于涉及实验动物生物科学研 究的任何领域。 	 该指南提供一个核对清单便于对准备 发表的稿件进行准备和评审。 参考文献 1. Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG (2010) Improving Bioscience Research Reporting: The ARRIVE Guidelines for Reporting Animal Research. PLOS Biol 8(6): e1000412. doi:10.1371/journal.pbio.1000412 2. Schulz KF, Altman DG, Moher D, the CONSORT Group (2010) CONSORT 2010 Statement: updated guidelines for reporting parallel group randomised 	国家3R中心 (NC3Rs) 对所有在 ARRIVE 指南的创设中提供宝贵 经验和建议的各位专家致以最衷 心的感谢。特别感谢NC3Rs 报告 指南工作组所有成员。我们同时要 感谢NC3Rs 的基金持有者、医学 研究委员会、生物技术和生物科学 研究委员、威康信托、帕金森氏病 协会、英国心脏基金会,及其基金 持有人和基金委员会成员对指南 所提的反馈意见。 感谢胡晖编辑(《环境与健康展 望》)、庞万勇博士(赛诺菲研发中
	ARRIVE指南的适用对象是: • 初涉写作或经验丰富的作者 • 杂志编辑 • 专业评审 • 资助机构		trials. BMJ 340:c332. 资助 ARRIVE 报告指南项目是由国家3R (替代,优化,和减少使用动物进行研 究) 中心 (NC3Rs) 资助	 心)和美东开博士(化教大学均附 女王学院)帮助审核中文翻译。 Further Information www.nc3rs.org.uk/ARRIVE enquiries@nc3rs.org.uk

So we would find more 3R literature if there was greater transparency...

Improved publication standards Open Access to primary data and negative results Clear implementation of the 3Rs Editorial action

- More structured M&M sections in papers
- Information on the ethical review process and justification
- Experimental design and appropriate analysis
- Compliance with guidelines
 - ARRIVE, GSPC, ILAR, ICLAS, LASA/APC
 - local guidelines, AAALAC template http://oslovet.norecopa.no/3R/produkter.aspx?search=reporting
- Compliance with the Basel Declaration



basel-declaration.org

- emphasises the 3Rs
- encourages transparency and collaboration to avoid repetition of animal studies
- implement and monitor the highest training standards
- invites animal welfare organisations to open discussion
- promotes balanced dialogue

We would also find more 3R literature if there was greater use of 3R descriptors...

Using PubMed to access data in MEDLINE:

MESH (Medical Subject Headings) thesaurus

PubMed "Animal Testing Alternatives"[Me
Summary - 20 per page - Sort by Most Recent -
Results: 1 to 20 of 2635
Animal Experimentation"[Mesh]
RSS Save search Advanced
per page - Sort by Most Recent -

Other databases have their own thesauri. A thesaurus can be useful to build up a list of suitable keywords, even if you use another database.

Examples of 3R sources

- National 3R centres
- 3R congress proceedings
- Guidelines papers
- Journals
- Discussion groups
- Training schools

National 3Rs Centres



www.nc3rs.org.uk

Canadian Council on Animal Care (CCAC)

Guidelines for lab, farm, fish and wildlife research

╋

HOME Send us your feedback Three Rs Microsite CCAC web site

Step-by-step Three Rs search strategy

Quick Info

CCAC guidelines & policies on animal care protocols

Where to do a Three Rs literature search

is your Three Rs Search complete?

Animal use protocol worksheet

www.ccac.ca

Three Rs Search Guide

If you plan to use animals for scientific purposes, you must complete an animal use protocol and submit it animal to an care committee for approval prior to commencement of the The animal study. use protocol outlines how the Three Rs will be the implemented in proposed animal-based procedures. To find the most up-to-date information Three the Rs, on investigators typically conduct а structured information search. To assist investigators with this search, the CCAC has Rs produced the Three Search Guide.



The Three Rs Search Guide provides detailed instructions on how to conduct a Three Rs information search in the <u>Step-by-Step Three Rs</u> <u>Search Strategy</u>.

Animal welfare organisations



My RSPCA Help

UFAW/RSPCA Rabbit Behaviour and Welfare Group

During 2008, the UFAW/RSPCA Rabbit Behaviour and Welfare Group published a report providing practical guidance on refining laboratory rabbit husbandry

Reducing suffering:

For as long as animals are used in research and testing, every step must be taken to reduce suffering and

AND ADVICE 0300 1234

The research animals department promotes initiatives that will lead to improvements in laboratory animal housing and care and reductions in suffering caused by procedures ...

Working to improve the welfare of laboratory rodents is extremely important because the vast majority of animals used in research and testing



The lives of laboratory rabbits can be greatly improved by providing housing and care that caters for their physical and behavioural needs more

www.rspca.org.uk/sciencegroup/researchanimals



A resource book for lay members of ethical review and similar bodies worldwide

3rd edition January 2015

Maggy Jennings and Jane A. Smith



Centres giving information on alternatives





University of California Center for Animal Alternatives



www.lib.ucdavis.edu/dept/animalalternatives



Animal Welfare Information Center

U.S. DEPARTMENT OF AGRICULTURE NATIONAL AGRICULTURAL LIBRARY

awic.nal.usda.gov

Future collaboration: retrieval of specific, flagged 3R-records from a Unified Repository



Animal Welfare Information

Center

U.S. DEPARTMENT OF AGRICULTURE NATIONAL AGRICULTURAL LIBRARY

accessing references from many of the large databases such as MEDLINE

norecopa.no awic.nal.usda.gov

The world congresses on the 3Rs are important 3Rdrivers and disseminators of information: wc9prague.org

891 abstracts, 49 countries, 1000 participants (the next one is in September 2017 in Seattle)

1996: 2nd World Congress on Alternatives and Animal Use in the Life Sciences, Utrecht



1997: Altweb (Alternatives to animals on the web) http://altweb.jhsph.edu

International consensus meetings

Harmonisation of the Care and Use of: Fish (2005) Wildlife (2008) Fish (2009) Agricultural animals (2012)

http://norecopa.no/consensus-meetings

All presentations and consensus statements are on the internet: a lasting resource

Guidelines as a portal to more information

R Johansen, JR Needham, DJ Colquhoun, TT Poppe & AJ Smith

Guidelines for health and welfare monitoring of fish used in research

Laboratory Animals, 2006, 40: 323-340 http://www.lal.org.uk/pdffiles/GuidelinesFish.pdf

Search 3R Guide 0 Find 3R resources ✓ - All Categories -Agricultural animals Anaesthesia and analgesia Aquatic animals Behavioural research Birds Blood sampling Cancer research Design Disease research Education and training Environmental enrichment Ethics & harm-benefit analysis Fish Handling Housing and management Humane killing Miscellaneous Neuroscience research Non-human primates Nutritional research Procedures Reporting Surgical research Toxicology Transport Wildlife

For a global view of guidelines, see 3R Guide:

http://oslovet.norecopa.no/3R/produkter.aspx?type=66_Guidelines



Expert Working Group report on severity classification

Sundached in support of the revision of Literative 55-009/3235, on the protection of manuals used is classific purposes

Conducted in support of the revision of Directive 86/609/EEC on the protection of animals used for scientific purposes

Expert working group on severity classification of scientific procedures performed on animals

FINAL REPORT Brussels, July 2009

http://ec.europa.eu/environment/chemicals/lab_animals/pdf/report_ewg.pdf

Methods of positioning fish for surgery or other procedures out of water

Trond Brattelid & Adrian J. Smith

Laboratory Animal Unit, Norwegian School of Veterinary Science, PO Box 8146 Dep., N-0033 Oslo, Norway



We need more guidelines for specific research areas

Published online on 9 May 2011 Lab Anim, doi: 10.1258/a.2011.01018

Working Party Report

Guidance on the severity classification of scientific procedures involving fish: report of a Working Group appointed by the Norwegian Consensus-Platform for the Replacement, Reduction and Refinement of animal experiments (Norecopa)

P Hawkins (Convenor)¹, N Dennison², G Goodman³, S Hetherington⁴, S Llywelyn-Jones⁵, K Ryder² and A J Smith⁶

Research Animals Department, RSPCA, Wilberforce Way, Southwater, West Sussex RHI3 9RS, UK, ⁵Animals (Scientific Procedures Inspectorate, Home Office, PO Box 6779, Dundee DD1 9WW, UK: *Bological Services, The University of Edinburgh, Charcelor Building, 49, Little France Crescent, Edinburgh EH16 45B, UK, "DEFAS, Palefield Road, Lowestoft, NR33 0HT, UK; "Ning's College London, Biological Services Unit, 4th Soy, Hodnide Building, Guar's Campus, London SE1 1UL, UK, "Noranoos, e./o. Norwenian Vaterham Institute, PO Box 750 Sentrum, N-0105 Oslo, Norway Corresponding author: P Hawkins, Email: phawkins@rspca.org.uk

Abstract

The severity classification of procedures using animals is an important tool to help focus the implementation of refinement and to assist in sporting the application of the 3Rs (splacement, induction and refinement). The recently revised Directive that regulates animal measurch and testing within the European Union moving Member States to ensure that all opportunes are classified as 'non-recovery', 'mild', 'moderate' or 'severs', using assignment offeria set out by the European Commission (EC). However, these are focused upon terrestrial species, so are of limited relevance to fish users. A Working Group set up by the Norwegian Consensus-Platform for the 3Rs (Norecopa) has produced guidance on the classification of severity in scientific procedures involving fish, including examples of "subthreshold", "mild", "moderate", "severe" and "upper threshold" procedures. The aims are to complement the EC guidelines and help to ensure that suffering infish is effectively predicted and ninimized. Norecopa has established a website (www.norecopa.no/categories) where more information on severity classification for procedures using fish, including field research, will be made available.

Keywords: Fish, harm-benefit assessment, humane endopints, refinement, severith

Laboratory Animak 2011: 1-6. DOI: 10.1258/la.2011.01018

Background

An effective prediction of the effects of a research protocol on the animals concerned helps to ensure that any rain, sufforing or distress they may experience will be effectively anticipated, recognized and alloviated. This is essential not only for animal welfase but also for scientific validity, because physiological and behavioural responses to suffering can significantly affect data quality. Severity classifiimplementation of refinement, induding monitoring its pro-gress, and to assist in reporting the application of the 3Rs (replacement, reduction and refinement) of Russell and Burch.1 which is now an integral part of the legislation on animal research and testing in many countries. Predictions

assessments undertaken by bodies such as negulatory auth orities and ethical committees when deciding whether or not a project should be licensed or funded.

There may also be a legal requirement to predict and classity severity. For example, the new Directive regulating animal use within the European Union, which must be implemented within all Member States by January 2013, requires the severity of each procedure to be classified on the basis of the degree of pain, suffering, distress or cation is thus an important tool to help focus the lasting harm expected to be experienced by an individual animal during the course of the procedure, with the aim of enhancing transparency, facilitating the project authorization process and providing tools for monitoring compli-ance.² Member States will have to ensure that all very', 'mild', 'moder procedures are classified as 'non-record of severity are also fundamental to the harm-benefit ate or 'severe' on a case-by-case basis, using the assignment

> Laboratory Animals 2011: 1-8 Copyright 2011 by the Laboratory Animals Limited

P Hawkins, N Dennison, G Goodman, S Hetherington, S Llywelyn-Jones, K Ryder and AJ Smith

> Laboratory Animals, 45: 219-224, 2011 www.norecopa.no/categories

Guidance on the severity classification of procedures involving fish

Position Statements and Guidelines

- Food deprivation •
- Toe clipping •
- Pain relief •
- Fin clipping of fish •
- Biometric methods of • identification
- Methods for identification • of birds



Om Norecopa

Ofte stilte spørsmål

Styre og sekretariat Vedtekter

Styrets intranett

Faglige uttalelser

Konsensusmøter

Norecopas 3R-pris

Hva giør vi?

Aktivitetsplan

Årsmøter

Regnskap Styrereferater

Bli medlem!

Medlemmer

Nyhetsbrev

Activities

Medlemsfordeler

Tegn medlemskap!

English section

About Norecopa

3Rs resources

Guidelines A-Z

Categories of severity

Consensus meetings

Position statements

The Concept of the

of audiovisual

The NORINA database

alternatives in teaching and training Dyr i forskning Å planlegge dyreforsøk

3Rs

Fick Husdyr Laboratoriedyr

Textbase: literature on lab animal science

Statistical design

Historikk Informasjonsmateriell

norecopa

n > 3Rs resources > Position statements

Position statements

Norecopa produces position statements on topics related to the use of animals in research and the 3Rs.

Toe clipping in mice

The Norwegian Animal Research Authority asked Norecopa to evaluate toe clipping as a means of identification and tissue sampling in mice. The Board produced an 18-page document, which has been circulated to all members. A translation of the final version can be downloaded here. The document includes an evaluation of alternative methods for the identification and genotyping of rodents, with a literature references. The Board composed a supplementary statement in March 2010 following three new published studies, which was circulated to its members.

Pain relief in rodents

In collaboration with colleagues in the laboratory animal environment, Norecopa has produced a document on pain relief in rodents.

Food deprivation in rodents

Norecopa has written an 11-page position statement on food deprivation in rodents. The summary (recommendations) can be read here. The full document is available here.

Student essays

Norecopa has edited essays on Fin clipping in fish, Biometric methods of

identification and Guides to identification methods for birds, in connection with a course in laboratory animal science for researchers. These essays have not been qualitycontrolled, nor is the content necessarily endorsed by Norecopa. Updated versions of the essays will be published here, if we receive feedback from readers. The contents of these essays may be used freely, but it must not be presented as representing the views of Norecopa or its secretary. Furthermore, the factual content (including literature references) should be checked before use.

Would you like Norecopa to write a statement about a topic related to your research? Please contact the secretary!

Norecopa also produces political statements and writes newspaper articles about animal research and the 3Rs. These can be read here.



Om Norecona

Norecopa arbeider for å fremme "de 3 R'ene" i forskningen som kan involvere dyr:

* Replace * Reduce * Refine

Norecona tilstreber konsensus om de tre R'ene mellom alle de fire interessepartene rundt dyreforsøk:





Norecopa Norwegian Veterinary Institute P.O.Box 750 Sentrum N-0106 Oslo, Norway

Visiting address: Ullevälsveien 68 0454 Oslo Org.no. 992 199 199 Tel: +47 41 22 09 49 Fax: +47 23 21 60 01 post@norecopa.no www.norecona.no

Utviklet av Netlab Oppdateres med

Position Statements from the Norwegian Animal Research Authority



FORSØKSDYRUTVALGET

	 [03.07.14] Prinsipputtalelse om bruk av telemetri [03.07.14] Prinsipputtalelse om bruk av telemetri-halsbånd på hjortevilt og rovvilt (25.10.13] Søknadsplikt ved blodprøvetaking av viltlevende fugler (03.09.13] Prinsipputtalelse: Merking av viltlevende fugler (29.11.12) Prinsipputtalelse om blokksøknader (23.10.12) Merkemetoder på fisk i laboratorieforsøk (28.09.11) Krav til avdelinger som ønsker å bli godkjente forsøksdyrvirksomheter
Hiem	(18.03.11) Retningslinjer for behandling av søknader om forsøk med lakselus
Arkiv	[27.10.10] Alle søknader som involverer lakselus skal inntil videre behandles av FDU
Om forsøksdyrutvalget	(03.04.09) Smitteforsøk og smertevoldende forsøk på fisk
Regelverk	[14.11.08] Forsøksdyr med ein avvikande fenotype (genmodifiserte dyr, mutanter og innavla linjer)
Prinsippavgjørelser	(24.05.06) Utviklingsstadium for fiskelarver som omfattes av regelverk for forsøksdyr.
For søkere	102.06.04) Aviiving av gnagere med CO2 102.06.04) Giftighetstesting på fisk i petroleumsvirksomheten
Kurs og kompetansekrav	[26.08.03] Utfasing av LD50 - akutt giftighets testing
Ofte stilte spørsmål	[21.07.03] Forsøk med dyr for å illustrere kjent kunnskap (inkl. undervisning)
Alternativ til forsøk med dyr	[21.07.03] Veiledning for beredskapsvakt utenom ordinær arbeidstid ved Forsøksdyravdelinger [11.12.02] Bruk av eter til bedøvelse
Lenker	(11.12.02) Forsøksdyrutvalgets policy ved smertevoldende dyreforsøk
Møtevirksomhet	(03.07.02) Bruk av intraperitoneale radiosendere
Kontakt oss	[10.05.02] Avlivningsmetoder for nyfødte smågnagere

http://www.mattilsynet.no/fdu/prinsippavgjorelser

ec.europa.eu/animals-in-science



European Directive, Article 47: 3*R*-alternative approaches

- 1. The Commission and Member States shall contribute to the development and validation of 3R-alternatives, and encourage research in this field
- 2. Member States shall assist the Commission in identifying laboratories for validation studies
- 3. The Commission shall set the priorities for these studies and allocate tasks
- 4. Member States shall promote alternatives and disseminate information on them
- Member States shall nominate a single point of contact to provide advice on the regulatory relevance and suitability of alternatives proposed for validation (PARERE: Preliminary Assessment of Regulatory Relevance)

Journals

ATLA (Alternatives to Laboratory Animals) Animal Welfare (UFAW) ILAR Journal Laboratory Animals Comparative Medicine

See www.3RGuide.info for more

It doesn't have to be the latest issue or most recent report...



http://ilarjournal.oxfordjournals.org

Email discussion groups

e.g. CompMed + archive LAREF VOLE Local competent persons...

See www.3RGuide.info for more

FRAME Training Schools

Portugal, 30 March-1 April 2015 Norway, February 2016

www.frame.org.uk/training-schools



NC3Rs website

http://nc3rs.org.uk/experimental-design



National Centre for the Replacement Refinement & Reduction of Animals in Research Guidelines for the Design and Statistical Analysis of Experiments Using Laboratory Animals http://ilarjournal.oxfordjournals.org/content/43/4/244.full

NC3Rs Experimental Design Assistant (EDA) http://nc3rs.org.uk/experimental-design-assistant-eda



TextBase

TextBase publications

Your search for TextBase publications containing the text "design" in the title returned the following results (13 items, page 1 of 1);

The Design of Animal Experiments: Reducing the Use of Animals in Research Through Better Experimental Design. By Festing, Michael F.W.; Overend, Philip, Das, Rose Gaines; Borja, Mario Cortina & Berdoy, Manuel (2002). This handbook is aimed at all research scientists who use laboratory animals, with the aim of helping them to design their own experiments more effectively and/or to improve their ability to communicate with professional statisticians when designing more complex experiments.

CCAC Guidelines on: Laboratory Animal Facilities -

Characteristics, Design, and Development. By Neil, David and McKay, Donald, with the collaboration of the CCAC Facilities Standards Subcommittees (2003). This document concentrates on the characteristics of a laboratory animal facility and hence do not cover all subjects matter discussed in the "Guide to the Care and Use of Experimental Animals", Volume 1, Chapters II and III, (CCAC, 1993).

Experimental Design and Analysis in Animal Sciences. By Morris, Tim R. (1999). This guide includes information for the design and analysis of experiment in animal science.

Experimental Design: A Handbook and Dictionary for Medical and Behavioral Research. By Krauth, J. (2000). Scientists planning experiments in medical and behavioural research will find this handbook and dictionary an invaluable desk reference tool.

oslovet.norecopa.no/textbase



Entire website

+

http://www.uk.sagepub.com/books/Book242188? siteId=sage-uk&prodTypes=any&q=9781853155130