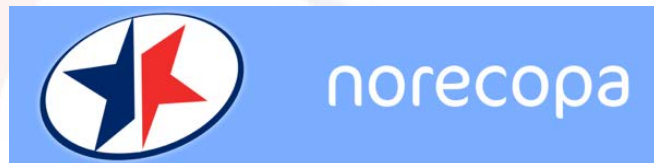


**PREPARE before you ARRIVE:
Good reporting relies on good planning**

norecopa.no/prepareFELASA

Adrian Smith

adrian.smith@norecopa.no



<https://norecopa.no>



*‘Our long experience and modern coffee machines
are your guarantee of quality’*

Scientists are becoming increasingly concerned about the validity of animal experiments

NATURE | NEWS

Swiss survey highlights potential flaws in animal studies

Poor experimental design and statistical analysis could contribute to widespread problems in reproducing preclinical animal experiments

Pain management in pigs undergoing experimental surgery; a literature review (2012–4) FREE

A. G. Bradbury, M. Eddleston, R. E. Clutton

Br J Anaesth (2016) 116 (1): 37-45. DOI: <https://doi.org/10.1093/bja/aev301>

Published: 03 October 2015

selection criteria. Most articles (193/233, 83%) described use of drugs with analgesic properties, but only 87/233 (37%) described postoperative analgesia. No article provided justification for the analgesic chosen, despite the lack of guidelines for analgesia in porcine surgical models and the lack of formal studies on this subject. Postoperative pain assessment was reported in only 23/233 (10%) articles. It was found that the reporting of postoperative pain management in the studies was remarkably low, reflecting either under-reporting or under-use. Analgesic description, when given, was frequently too limited to enable reproducibility. Development of a

Norecopa: PREPARE for better research



NATURE | NEWS FEATURE

1,500 scientists lift the lid on reproducibility

Survey sheds light on the 'crisis' rocking research.

Monya Baker

25 May 2016 | Corrected: 28 July 2016

More than 70% of researchers have tried and failed to reproduce another scientist's experiments, and more than half have failed to reproduce their own experiments. Those are some of the telling figures that emerged from *Nature's* survey of 1,576 researchers who took a brief online questionnaire on reproducibility in research.

FELASA, 10-13 June 2019

How do they do it?



<https://www.meonuk.com/runway-markings-explained>



norecopa



Norecopa: PREPARE for better research

FELASA, 10-13 June 2019



norecopa



Norecopa: PREPARE for better research

FELASA, 10-13 June 2019



norecopa



kathmandupost.ekantipur.com



Norecopa: PREPARE for better research

FELASA, 10-13 June 2019

Aviation and Animal Research: Human Factors



A Pilot's Perspective
By Jake Hannabuss

Accident Rate for commercial flights is
one fatal accident per 16 million flights



Tools – Crew Briefing

- Division of **Responsibilities**
- Planned **Sequence of Events**, including **deviations from normal procedures**
- The **Routine Factors** to be considered
- Actions in the event of an **emergency**
- **Special considerations**, weather, terrain, abnormalities
- Pre take-off and pre-landing **briefings**

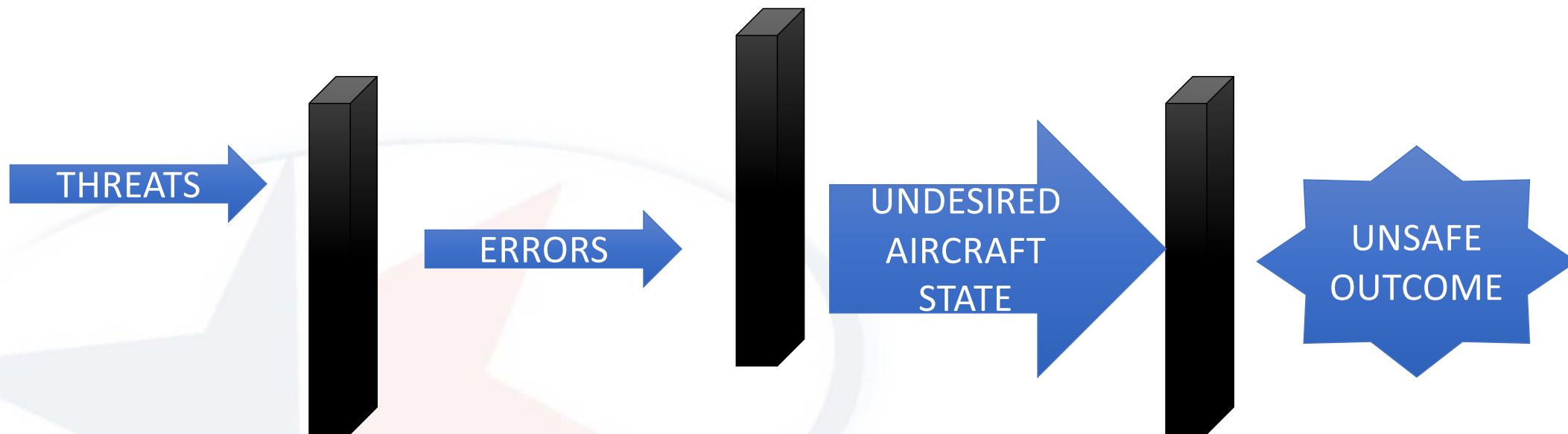
Checklists

- Reduce risk of **forgetting** to carry out vital actions
- Ensure checks are carried out in the **correct sequence**
- Encourage **cooperation** and **cross-checking** between crew members

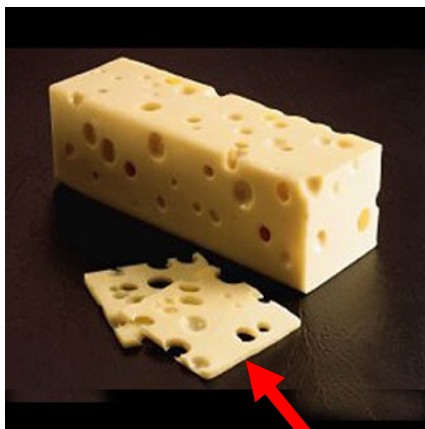
10-15 checklists on short European flights



- Threat and Error Management (TEM)
- Identifies a ***chain***, which precedes all ***unsafe outcomes***:



<https://airfactsjournal.com/2012/10/threat-and-error-management-a-primer>



eaugallecheese.com/Swiss-Cheese



"Layer of defence"
or redundancy

Weakness / hazard

Loss

wikipedia.org/wiki/Swiss_cheese_model



norecopa



Norecopa: PREPARE for better research

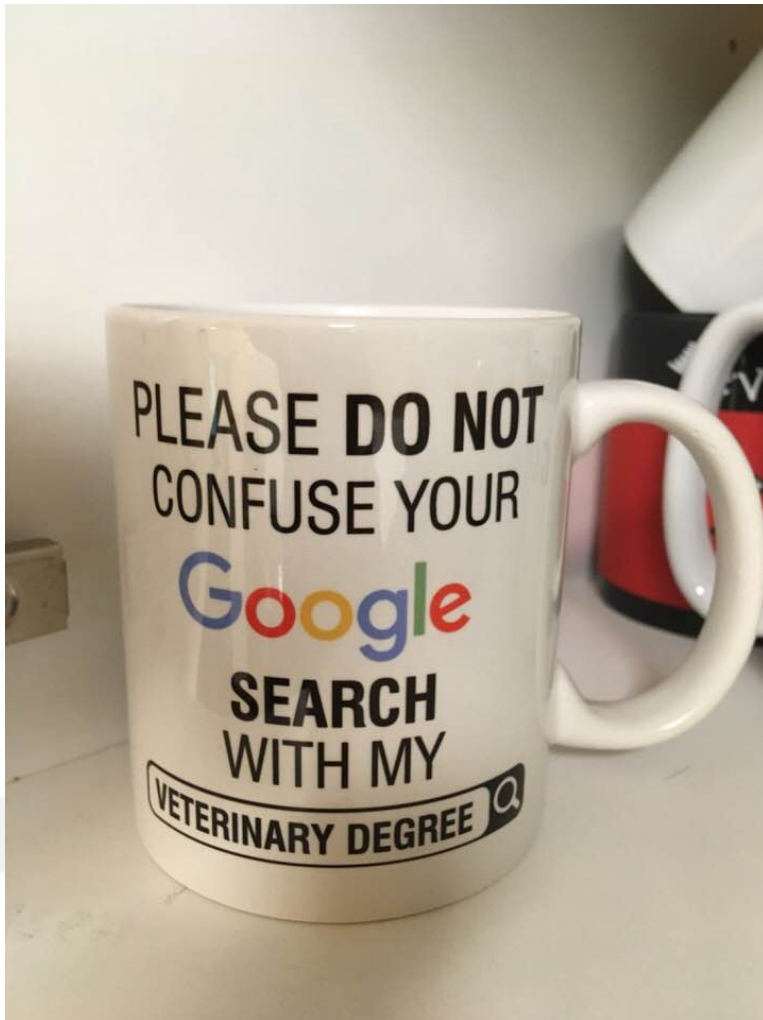
FELASA, 10-13 June 2019

How do we do it...?

Literature searching is so much more than Systematic Reviews...



norecopa



Norecopa: PREPARE for better research

How to construct a literature search

Alice Tillema, Medical Library, Nijmegen

<http://libguides.ru.nl/norecopa>



Radboud University



Radboudumc
university medical center

norecopa.no/more-resources/literature-searches-and-systematic-reviews

FELASA, 10-13 June 2019

Identification and elimination of contingent suffering



animalcaresystems.com

(not just the direct suffering caused by the procedure)

Fear, boredom and discomfort

Caused by, for example:

Transport, or changes in housing, husbandry and social groups

Single-housed male mice show symptoms of what in humans would be characterised as depression

<http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0111065>



photo: colourbox.com

Stress caused by capture and handling



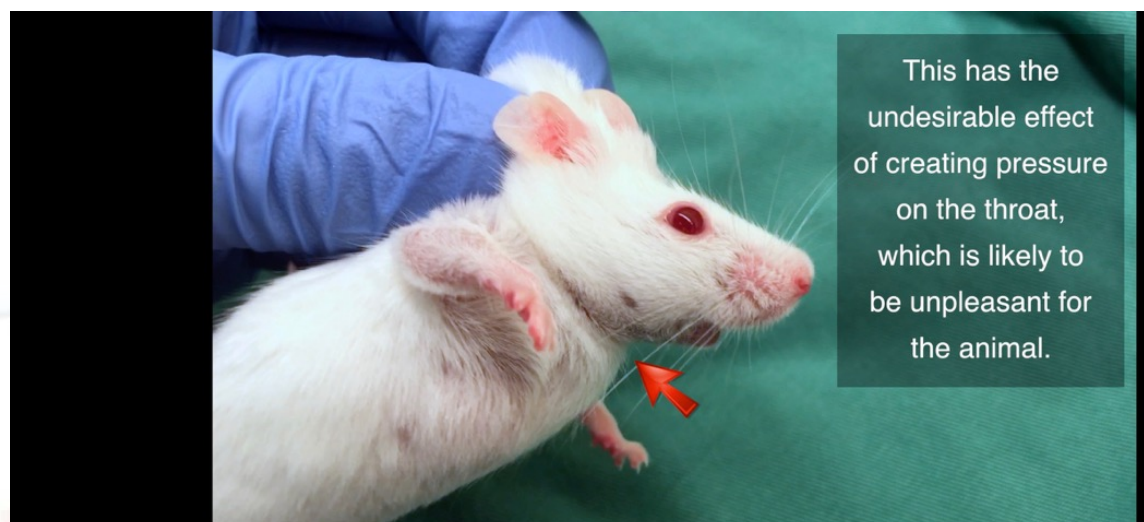
News > Science

Scores of scientific studies based on mice thrown into doubt because they were picked up by the tail

Mice picked up by the tail – standard practice in labs – are stressed and anxious so don't act naturally in some experiments, new study finds

Ian Johnston Science Correspondent | [@montaukian](#) | Tuesday 21 March 2017 10:58 GMT | [3 comments](#)

Stress caused by capture and handling



Three fingers better than two MATURE

5 days ago | More



▶ 1,016 ♥ 0 📁 0 💬 0

↓ Download ↩ Share

<http://bitly.com/scruff-technique>

More from Norecopa

Autoplay next video



Three fingers b
Norecopa



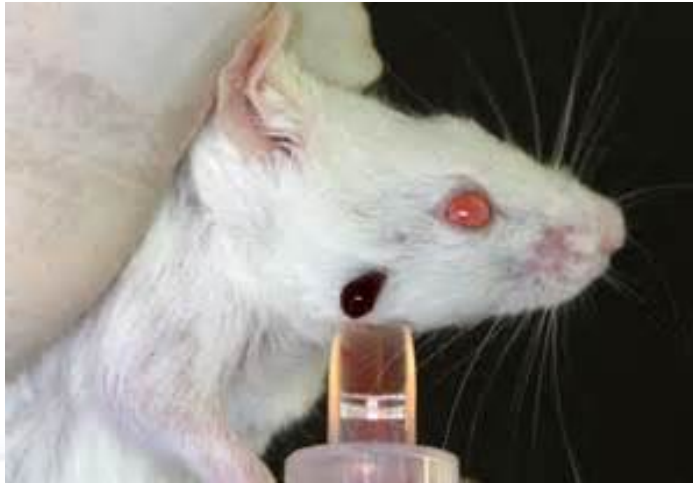
Artefacts caused by poor administration techniques



Photo: NMBU

- *Are you sure that your injection ends up in the same place each time?*
- *Are the injections painful?*
- *Are they realistic? (intramuscular injections in small animals)*

'Simple' blood sampling techniques?



medipoint.com/html/for_use_on_mice.html



The best blood sampling techniques are those where you can:

- ✓ see the blood vessel
- ✓ regulate the amount of blood you remove
- ✓ stop the bleeding easily and
- ✓ not damage the surrounding tissue

Carol M. Newton (1925-2014)



National Library of Medicine

The three S's

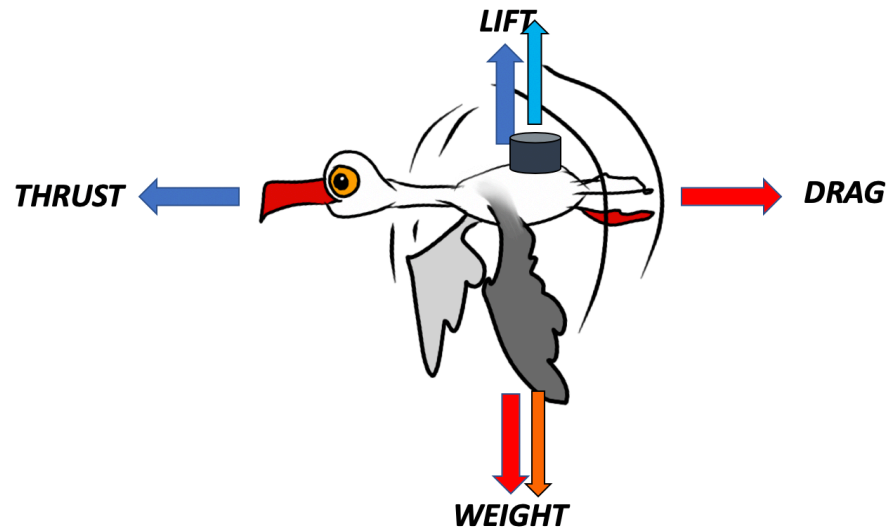
- *Good Science*
- *Good Sense*
- *Good Sensibilities*

norecopa.no/3S

Carol M Newton, quoted in Rowsell HC (1977): *The Ethics of Biomedical Experimentation in The Future of Animals, Cells, Models, and Systems in Research, Development, Education, and Testing* pp. 267-281, National Academy of Sciences, Washington, D.C., ISBN 0-309-02603-2.

Primary effects

The increased lift balances the extra force from the tag weight



From Rory Wilson: norecopa.no/media/8018/rory-wilson.pdf

Drag occurs in water as well as in the air...



From Rory Wilson: norecopa.no/media/8018/rory-wilson.pdf

Too late to read the checklists when you have arrived!



Norecopa: PREPARE for better research

colourbox.com

FELASA, 10-13 June 2019

Hurni 1969

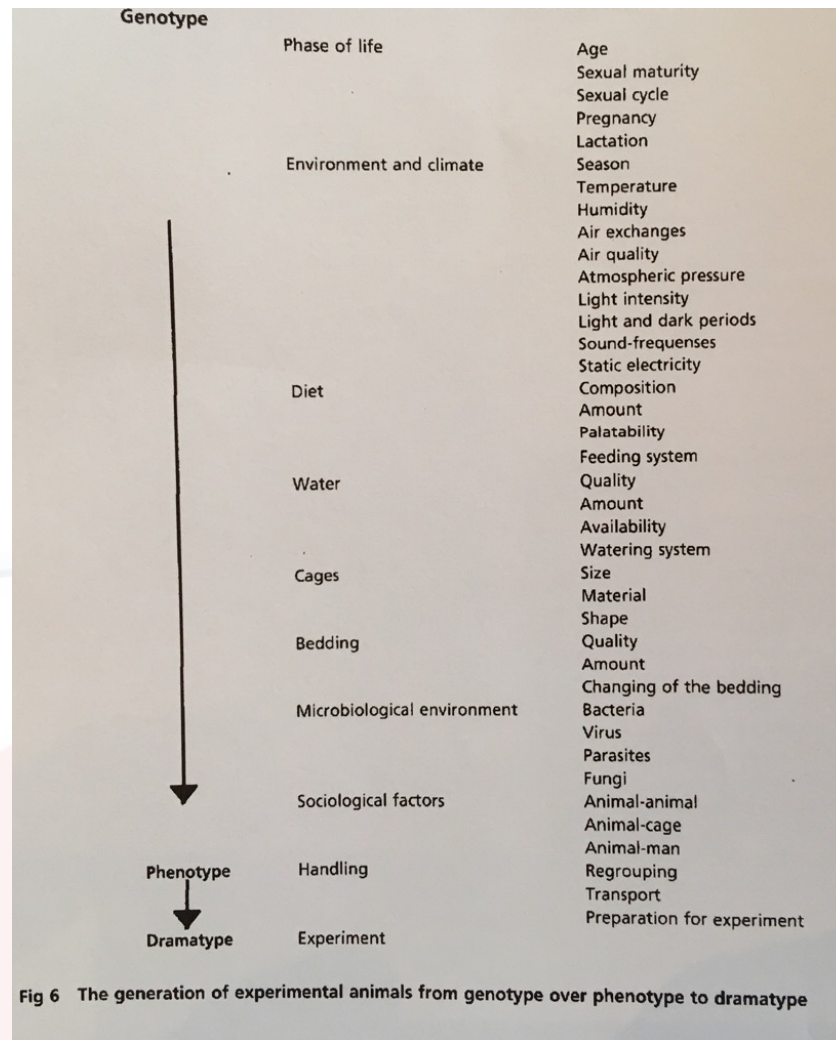


Fig 6 The generation of experimental animals from genotype over phenotype to dramatype

There are many guidelines for *reporting* animal studies

- Öbrink & Waller, 1996
- Reporting animal use in scientific papers (Jane Smith *et al.*), 1997
- Öbrink & Reh binder: Animal definition: a necessity for the validity of animal experiments? *Laboratory Animals*, 2000
- Guidelines for reporting the results of experiments on fish (2000)
- **ARRIVE Guidelines, 2010** (Kilkenny *et al.*, NC3Rs)
- **Gold Standard Publication Checklist, 2010 (SYRCLE)**
- Institute for Laboratory Animal Research, NRC, 2011
- Instructions to authors, in many journals
e.g. Nature's Reporting Checklist

The ARRIVE Guidelines



	ITEM	RECOMMENDATION
Title	1	Provide as accurate and concise a description of the content of the article as possible.
Abstract	2	Provide an accurate summary of the background, research objectives, including details of the species or strain of animal used, key methods, principal findings and conclusions of the study.
INTRODUCTION		
Background	3	<p>a. Include sufficient scientific background (including relevant references to previous work) to understand the motivation and context for the study, and explain the experimental approach and rationale.</p> <p>b. Explain how and why the animal species and model being used can address the scientific objectives and, where appropriate, the study's relevance to human biology.</p>
Objectives	4	Clearly describe the primary and any secondary objectives of the study, or specific hypotheses being tested.
METHODS		
Ethical statement	5	Indicate the nature of the ethical review permissions, relevant licences (e.g. Animal (Scientific Procedures) Act 1986), and national or institutional guidelines for the care and use of animals, that cover the research.
Study design	6	<p>For each experiment, give brief details of the study design including:</p> <p>a. The number of experimental and control groups.</p> <p>b. Any steps taken to minimise the effects of subjective bias when allocating animals to treatment (e.g. randomisation procedure) and when assessing results (e.g. if done, describe who was blinded and when).</p> <p>c. The experimental unit (e.g. a single animal, group or cage of animals).</p> <p>A time-line diagram or flow chart can be useful to illustrate how complex study designs were carried out.</p>
Experimental procedures	7	<p>For each experiment and each experimental group, including controls, provide precise details of all procedures carried out.</p> <p>For example:</p> <p>a. How (e.g. drug formulation and dose, site and route of administration, anaesthesia and analgesia used [including monitoring], surgical procedure, method of euthanasia). Provide details of any specialist equipment used, including suppliers).</p> <p>b. When (e.g. time of day).</p> <p>c. Where (e.g. home cage, laboratory, water maze).</p> <p>d. Why (e.g. rationale for choice of specific anaesthetic, route of administration, drug dose used).</p>
Experimental animals	8	<p>a. Provide details of the animals used, including species, strain, sex, developmental stage (e.g. mean or median age plus age range) and weight (e.g. mean or median weight plus weight range).</p> <p>b. Provide further relevant information such as the source of animals, international strain nomenclature, genetic modification status (e.g. knock-out or transgenic), genotype, health/immune status, drug or test naïve, previous procedures, etc.</p>

Housing and husbandry	9	<p>Provide details of:</p> <p>a. Housing (type of facility e.g. specific pathogen free (SPF); type of cage or housing; bedding material; number of cage companions; tank shape and material etc. for fish).</p> <p>b. Husbandry conditions (e.g. breeding programme, light/dark cycle, temperature, quality of water etc for fish, type of food, access to food and water, environmental enrichment).</p> <p>c. Welfare-related assessments and interventions that were carried out prior to, during, or after the experiment.</p>
Sample size	10	<p>a. Specify the total number of animals used in each experiment, and the number of animals in each experimental group.</p> <p>b. Explain how the number of animals was arrived at. Provide details of any sample size calculation used.</p> <p>c. Indicate the number of independent replications of each experiment, if relevant.</p>
Allocating animals to experimental groups	11	<p>a. Give full details of how animals were allocated to experimental groups, including randomisation or matching if done.</p> <p>b. Describe the order in which the animals in the different experimental groups were treated and assessed.</p>
Experimental outcomes	12	Clearly define the primary and secondary experimental outcomes assessed (e.g. cell death, molecular markers, behavioural changes).
Statistical methods	13	<p>a. Provide details of the statistical methods used for each analysis.</p> <p>b. Specify the unit of analysis for each dataset (e.g. single animal, group of animals, single neuron).</p> <p>c. Describe any methods used to assess whether the data met the assumptions of the statistical approach.</p>
RESULTS		
Baseline data	14	For each experimental group, report relevant characteristics and health status of animals (e.g. weight, microbiological status, and drug or test naïve) prior to treatment or testing (this information can often be tabulated).
Numbers analysed	15	<p>a. Report the number of animals in each group included in each analysis. Report absolute numbers (e.g. 10/20, not 50%).</p> <p>b. If any animals or data were not included in the analysis, explain why.</p>
Outcomes and estimation	16	Report the results for each analysis carried out, with a measure of precision (e.g. standard error or confidence interval).
Adverse events	17	<p>a. Give details of all important adverse events in each experimental group.</p> <p>b. Describe any modifications to the experimental protocols made to reduce adverse events.</p>
DISCUSSION		
Interpretation/scientific implications	18	<p>a. Interpret the results, taking into account the study objectives and hypotheses, current theory and other relevant studies in the literature.</p> <p>b. Comment on the study limitations including any potential sources of bias, any limitations of the animal model, and the imprecision associated with the results².</p> <p>c. Describe any implications of your experimental methods or findings for the (on the 3Rs) of the use of animals in research.</p> <p>Findings of this study are likely to translate to any relevance to human biology.</p> <p>(grant number) and the role of the funder(s)</p>



<https://www.nc3rs.org.uk/arrive-guidelines>

The ARRIVE guidelines

The ARRIVE guidelines claim that they ‘provide a logical checklist with **all the things that need to be considered when designing an experiment**’.

In our experience when planning animal research, **a number of additional points need to be addressed at the planning stage.**

These items improve

- **study quality**
- **animal welfare**
- and therefore **reproducibility**

- and also the **safety of humans and animals** affected directly or indirectly by the work



norecopa

ARRIVE

PREPARE

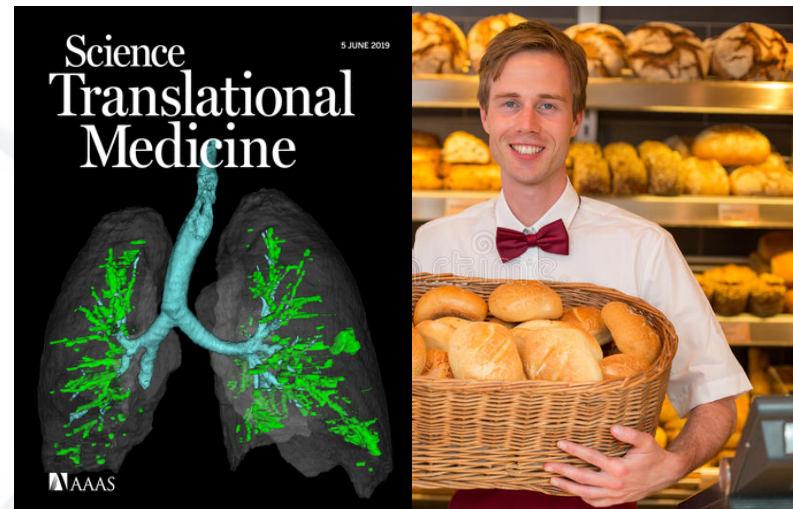
***Reporting guidelines like ARRIVE describe the experiment.
Guidelines like PREPARE are used to plan the experiment
(choose the «ingredients» and «baking time»)***



PREPARE *from day 1*

<https://www.bls.gov/ooh/images/3077.jpg>

ARRIVE



<https://www.dreamstime.com>



Original Article

PREPARE: guidelines for planning animal research and testing

Adrian J Smith¹, R Eddie Clutton², Elliot Lilley³, Kristine E Aa Hansen⁴ and Trond Bratteli⁵

Abstract
There is widespread concern about the quality, reproducibility and translatability of studies involving research animals. Although there are a number of reporting guidelines available, there is very little overarching guidance on how to *plan* animal experiments, despite the fact that this is the logical place to start ensuring quality. In this paper we present the PREPARE guidelines: Planning Research and Experimental Procedures on Animals: Recommendations for Excellence. PREPARE covers the three broad areas which determine the quality of the preparation for animal studies: formulation, dialogue between scientists and the animal facility, and quality control of the various components in the study. Some topics overlap and the PREPARE checklist should be adapted to suit specific needs, for example in field research. Advice on use of the checklist is available on the Norecopa website, with links to guidelines for animal research and testing, at <https://norecopa.no/PREPARE>.

Keywords
guidelines, planning, design, animal experiments, animal research

Date received: 5 April 2017; accepted: 27 June 2017

Introduction
The quality of animal-based studies is under increasing scrutiny, for good scientific and ethical reasons. Studies of papers reporting animal experiments have revealed alarming deficiencies in the information provided,^{1,2} even after the production and journal endorsement of reporting guidelines.³ There is also widespread concern about the lack of reproducibility and translatability of laboratory animal research.⁴⁻⁷ This can, for example, contribute towards the failure of drugs when they enter human trials.⁸ These issues come in addition to other concerns, not unique to animal research, about publication bias, which tends to favour the reporting of positive results and can lead to the acceptance of claims as fact.⁹ This has understandably sparked a demand for reduced waste when planning experiments involving animals.¹⁰⁻¹² Reporting guidelines alone cannot solve the problem of wasteful experimentation, but thorough planning will increase the likelihood of success and is an important step in the implementation of the 3Rs of Russell & Burch (replacement, reduction, refinement).¹³ The importance of attention to detail at all stages is,

in our experience, often underestimated by scientists. Even small practical details can cause omissions or artefacts that can ruin experiments which in all other respects have been well-designed, and generate health risks for all involved. There is therefore, in our opinion, an urgent need for detailed but overarching guidelines for researchers on how to plan animal experiments which are safe and scientifically sound, address animal

Corresponding author:
Adrian Smith, Norecopa, c/o Norwegian Veterinary Institute, P.O. Box 750 Sentrum, 0104 Oslo, Norway.
Email: adrian.smith@norecopa.no

Laboratory Animals
0(0) 1-7
© The Author(s) 2017
Reprints and permissions:
sagepub.co.uk/journalsPermissions.nav
DOI: 10.1177/0023677217724823
journals.sagepub.com/home/lan
SAGE

Pre-published under Open Access on 3 August 2017,
sponsored by the Universities Federation for Animal
Welfare (UFAW), UK

<https://doi.org/10.1177/0023677217724823>



Over 8,000 downloads from the
journal website so far

Also downloadable from
norecopa.no/PREPARE

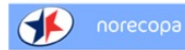
Norecopa: PREPARE for better research

FELASA, 10-13 June 2019

A downloadable checklist



PREPARE



The PREPARE Guidelines Checklist

Planning Research and Experimental Procedures on Animals: Recommendations for Excellence

Adrian J. Smith¹, R. Eddie Clutton², Elliot Lilley³, Kristine E. Aa. Hansen⁴ & Trond Brattelid⁵

¹Norecopa, c/o Norwegian Veterinary Institute, P.O. Box 750 Sentrum, 0106 Oslo, Norway; ²Royal (Dick) School of Veterinary Studies, Easter Bush, Midlothian, EH25 9RG, U.K.; ³Research Animals Department, Science Group, RSPCA, Wilberforce Way, Southwater, Horsham, West Sussex, RH13 9HS, U.K.; ⁴Section of Experimental Biomedicine, Department of Production Animal Clinical Sciences, Faculty of Veterinary Medicine, Norwegian University of Life Sciences, P.O. Box 8146 Dep., 0033 Oslo, Norway; ⁵Division for Research Management and External Funding, Western Norway University of Applied Sciences, 5020 Bergen, Norway.

PREPARE¹ consists of planning guidelines which are complementary to reporting guidelines such as ARRIVE². PREPARE covers the three broad areas which determine the quality of the preparation for animal studies:

1. Formulation of the study
2. Dialogue between scientists and the animal facility
3. Quality control of the components in the study

The topics will not always be addressed in the order in which they are presented here, and some topics overlap. The PREPARE checklist can be adapted to meet special needs, such as field studies. PREPARE includes guidance on the management of animal facilities, since in-house experiments are dependent upon their quality. The full version of the guidelines is available on the Norecopa website, with links to global resources, at <https://norecopa.no/PREPARE>.

The PREPARE guidelines are a dynamic set which will evolve as more species- and situation-specific guidelines are produced, and as best practice within Laboratory Animal Science progresses.

Topic	Recommendation
(A) Formulation of the study	
1. Literature searches	<input type="checkbox"/> Form a clear hypothesis, with primary and secondary outcomes. <input type="checkbox"/> Consider the use of systematic reviews. <input type="checkbox"/> Decide upon databases and information specialists to be consulted, and construct search terms. <input type="checkbox"/> Assess the relevance of the species to be used, its biology and suitability to answer the experimental questions with the least suffering, and its welfare needs. <input type="checkbox"/> Assess the reproducibility and transatability of the project.
2. Legal issues	<input type="checkbox"/> Consider how the research is affected by relevant legislation for animal research and other areas, e.g. animal transport, occupational health and safety. <input type="checkbox"/> Locate relevant guidance documents (e.g. EU guidance on project evaluation).
3. Ethical issues, harm-benefit assessment and humane endpoints	<input type="checkbox"/> Construct a lay summary. <input type="checkbox"/> In dialogue with ethics committees, consider whether statements about this type of research have already been produced. <input type="checkbox"/> Address the 3Rs (replacement, reduction, refinement) and the 3Ss (good science, good sense, good sensibilities). <input type="checkbox"/> Consider pre-registration and the publication of negative results. <input type="checkbox"/> Perform a harm-benefit assessment and justify any likely animal harm. <input type="checkbox"/> Discuss the learning objectives, if the animal use is for educational or training purposes. <input type="checkbox"/> Allocate a severity classification to the project. <input type="checkbox"/> Define objective, easily measurable and unequivocal humane endpoints. <input type="checkbox"/> Discuss the justification, if any, for death as an end-point.
4. Experimental design and statistical analysis	<input type="checkbox"/> Consider pilot studies, statistical power and significance levels. <input type="checkbox"/> Define the experimental unit and decide upon animal numbers. <input type="checkbox"/> Choose methods of randomisation, prevent observer bias, and decide upon inclusion and exclusion criteria.

Topic	Recommendation
(B) Dialogue between scientists and the animal facility	
5. Objectives and timescale, funding and division of labour	<input type="checkbox"/> Arrange meetings with all relevant staff when early plans for the project exist. <input type="checkbox"/> Construct an approximate timescale for the project, indicating the need for assistance with preparation, animal care, procedures and waste disposal/decontamination. <input type="checkbox"/> Discuss and disclose all expected and potential costs. <input type="checkbox"/> Construct a detailed plan for division of labour and expenses at all stages of the study.
6. Facility evaluation	<input type="checkbox"/> Conduct a physical inspection of the facilities, to evaluate building and equipment standards and needs. <input type="checkbox"/> Discuss staffing levels at times of extra risk.
7. Education and training	<input type="checkbox"/> Assess the current competence of staff members and the need for further education or training prior to the study.
8. Health risks, waste disposal and decontamination	<input type="checkbox"/> Perform a risk assessment, in collaboration with the animal facility, for all persons and animals affected directly or indirectly by the study. <input type="checkbox"/> Assess, and if necessary produce, specific guidance for all stages of the project. <input type="checkbox"/> Discuss means for containment, decontamination, and disposal of all items in the study.
(C) Quality control of the components in the study	
9. Test substances and procedures	<input type="checkbox"/> Provide as much information as possible about test substances. <input type="checkbox"/> Consider the feasibility and validity of test procedures and the skills needed to perform them.
10. Experimental animals	<input type="checkbox"/> Decide upon the characteristics of the animals that are essential for the study and for reporting. <input type="checkbox"/> Avoid generation of surplus animals.
11. Quarantine and health monitoring	<input type="checkbox"/> Discuss the animals' likely health status, any needs for transport, quarantine and isolation, health monitoring and consequences for the personnel.
12. Housing and husbandry	<input type="checkbox"/> Attend to the animals' specific instincts and needs, in collaboration with expert staff. <input type="checkbox"/> Discuss acclimatization, optimal housing conditions and procedures, environmental factors and any experimental limitations on these (e.g. food deprivation, solitary housing).
13. Experimental procedures	<input type="checkbox"/> Develop refined procedures for capture, immobilisation, marking, and release or rehoming. <input type="checkbox"/> Develop refined procedures for substance administration, sampling, sedation and anaesthesia, surgery and other techniques.
14. Humane killing, release, reuse or rehoming	<input type="checkbox"/> Consult relevant legislation and guidelines well in advance of the study. <input type="checkbox"/> Define primary and emergency methods for humane killing. <input type="checkbox"/> Assess the competence of those who may have to perform these tasks.
15. Necropsy	<input type="checkbox"/> Construct a systematic plan for all stages of necropsy, including location, and identification of all animals and samples.


References

1. Smith AJ, Clutton RE, Lilley E, Hansen KEA & Brattelid T. PREPARE: Guidelines for Planning Animal Research and Testing. *Laboratory Animals*, 2017, DOI: 10.1177/002367171724823.
2. Kilkenny C, Brown WJ, Cuthill IC et al. Improving Bioscience Research Reporting: The ARRIVE Guidelines for Reporting Animal Research. *PLoS Biology*, 2010, DOI: 10.1371/journal.pbio.1000412.

Further information

<https://norecopa.no/PREPARE> | post@norecopa.no | [@norecopa](#)



PREPARE 

The PREPARE Guidelines Checklist
'Planning Research and Experimental Procedures on Animals: Recommendations for Excellence'
 Adrian J. Smith, R. Eddie Clifton, Eilert Lütjens, Kristine G. Aa, Hansson & Trond Bratberg*

*Norecopa, c/o Norwegian Veterinary Institute, P.O. Box 750 Sentrum, 0166 Oslo, Norway; *Royal (Dick) School of Veterinary Studies, Easter Bush, Midlothian, EH25 9RG, U.K.; *Research Animals Department, Science Group, RSPCA, Wilberforce Way, Southwark, London, WC2R 2JF, U.K.; *Sector of Experimental Biomedicine, Department of Professor Arnekleiv Clinical Sciences, Faculty of Veterinary Medicine, Norwegian University of Life Sciences, P.O. Box 8148 Dep., 0203 Oslo, Norway; *Division for Research Management and External Funding, Western Norway University of Applied Sciences, 5020 Bergen, Norway.

PREPARE* consists of planning guidelines which are complementary to reporting guidelines such as ARRIVE.
 PREPARE covers the three broad areas which determine the quality of the preparation for animal studies:

1. Formulation of the study
2. Dialogue between scientists and the animal facility
3. Quality control of the components in the study

The topics will not always be addressed in the order in which they are presented here, and some topics overlap. The PREPARE checklist can be adapted to meet special needs, such as field studies. PREPARE includes guidance on the management of animal facilities, since in-house experiments are dependent upon their quality. The full version of the guidelines is available on the Norecopa website, with links to global resources, at <https://norecopa.no/prepare>.
 The PREPARE guidelines are a dynamic set which will evolve as more species- and situation-specific guidelines are produced, and as best practice within Laboratory Animal Science progresses.

Topic	Recommendation
(A) Formulation of the study	
1. Literature searches	<input type="checkbox"/> Form a clear hypothesis, with primary and secondary outcomes. <input type="checkbox"/> Consider the use of systematic reviews. <input type="checkbox"/> Decide upon databases and information specialists to be consulted, and construct search terms. <input type="checkbox"/> Assess the relevance of the species to be used, its biology and suitability to answer the experimental questions with the least suffering, and to welfare needs. <input type="checkbox"/> Assess the reproducibility and transferability of the project.
2. Legal issues	<input type="checkbox"/> Consider how the research is affected by relevant legislation for animal research and other areas, e.g. animal transport, occupational health and safety. <input type="checkbox"/> Locate relevant guidance documents (e.g. EU guidance on project evaluation).
3. Ethical issues, harm-benefit assessment and humane endpoints	<input type="checkbox"/> Construct a lay summary. <input type="checkbox"/> In dialogue with ethics committees, consider whether statements about this type of research have already been produced. <input type="checkbox"/> Address the 3Rs (replacement, reduction, refinement) and the 3Ss (good science, good sense, good sensibilities). <input type="checkbox"/> Consider pre-registration and the publication of negative results. <input type="checkbox"/> Perform a harm-benefit assessment and justify any likely animal harm. <input type="checkbox"/> Discuss the learning objectives, if the animal use is for educational or training purposes. <input type="checkbox"/> Allocate a severity classification to the project. <input type="checkbox"/> Define objective, easily measurable and unequivocal humane endpoints. <input type="checkbox"/> Discuss the justification, if any, for death as an end-point.
4. Experimental design and statistical analysis	<input type="checkbox"/> Consider pilot studies, statistical power and significance levels. <input type="checkbox"/> Define the experimental unit and decide upon animal numbers. <input type="checkbox"/> Choose methods of randomisation, prevent observer bias, and decide upon inclusion and exclusion criteria.

norecopa.no/prepare/prepare-checklist

PREPARE:

Planning **R**esearch and **E**xperimental **P**rocedures on **A**nimals: **R**ecommendations for **E**xcellence

PREPARE covers 15 topics:

Formulation of the study

1. Literature searches
2. Legal issues
3. Ethical issues, harm-benefit assessment and humane endpoints
4. Experimental design and statistical analysis

Dialogue between scientists and the animal facility

5. Objectives and timescale, funding and division of labour
6. Facility evaluation
7. Education and training
8. Health risks, waste disposal and decontamination

Methods

9. Test substances and procedures
10. Experimental animals
11. Quarantine and health monitoring
12. Housing and husbandry
13. Experimental procedures
14. Humane killing, release, reuse or rehoming
15. Necropsy

Items in pink are
not highlighted in
ARRIVE



In addition to the checklist, much more information is available on:

norecoba.no/PREPARE

A screenshot of the norecoba.no website. The header is blue with the Norecoba logo and the word "norecoba" in white. To the right, there are language options "NORSK" and "ENGLISH" with "ENGLISH" selected. Below the logo is a search bar with the text "Search:" and a magnifying glass icon. A navigation menu is located below the search bar, with the "PREPARE" link circled in red. The main content area lists various topics related to the PREPARE checklist, including literature searches, legal issues, ethical issues, experimental design, objectives, facility evaluation, education, health risks, test substances, experimental animals, quarantine, housing, experimental procedures, humane killing, necropsy, and comparison with ARRIVE. At the bottom of the page, there are social media icons for Facebook, Twitter, Email, and a plus sign for more options.



- PREPARE Checklist | 1-Literature searches | 2-Legal issues | 3-Ethical issues, Harm-Benefit Assessment and humane endpoints | 4-Experimental design and statistical analysis | 5-Objectives and timescale, funding and division of labour | 6-Facility evaluation | 7-Education and training | 8-Health risks, waste disposal and decontamination | 9-Test substances and procedures | 10-Experimental animals | 11-Quarantine and health monitoring | 12-Housing and husbandry | 13-Experimental procedures | 14-Humane killing, release, re-use or re-homing | 15-Necropsy | Comparison with ARRIVE

norecopa.no / PREPARE

Harm-Benefit Assessment

Harm-Benefit assessment, an evaluation of the likely sources and level of suffering of a planned procedure, followed by an assessment of the potential benefits of the research weighed against these harms, lies at the heart of [legislation in the EU](#) and elsewhere. [A framework for severity assessment and severity classification](#) must be established and justified. The likely adverse effects of each procedure should be described, along with their likely incidence and methods of recognising them, with indications of how these effects can be mitigated by implementing refinement. This necessitates the involvement of personnel with the relevant expertise to recognise, assess and reduce animal suffering, especially severe suffering. [Guidance on this is available on the RSPCA website](#). Specific justification of all unavoidable animal suffering must be provided. An estimate must be made of the maximum amount of pain, distress or lasting harm to which an individual can be

Links to quality guidelines worldwide on e.g. blood sampling, injection volumes, housing and husbandry, analgesia, humane endpoints, experimental design

Contract between the animal facility and the research group

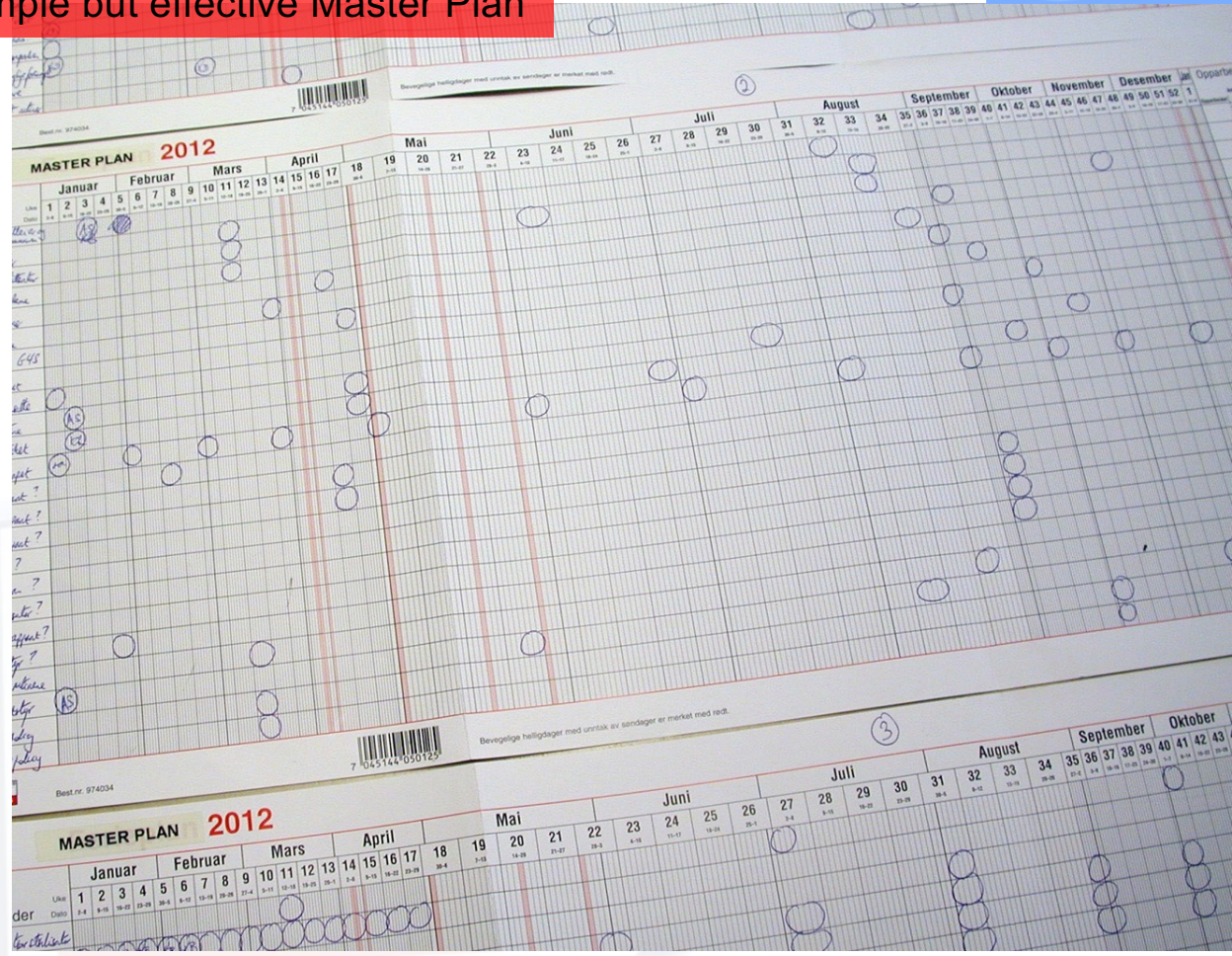
The division of labour and responsibilities

Clarifying all stages of the experiment

Ensuring that all necessary parameters are recorded

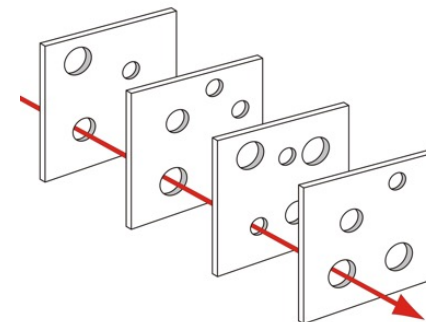
	Animal facility	Researcher	Not applicable
Animal:			
Arrival date			
Species			
Strain/stock and substrain			
Supplier (full name and address) or bred on the premises			
Number and sex			
Age, weight, stage of life cycle on arrival			
Pre-treatment (surgical or medical) from supplier			
Quality (e.g. SPF, germ-free, gnotobiotic, conventional)			
Acclimation time before the start of the experiment			
Time and duration of fasting (with/without water and bedding)			
Environment:			
Type of housing: barrier/conventional			
Temperature (mean ± variation)			
Light schedule			
Relative humidity (mean ± variation)			
Number of air changes in the animal room/cabinet per hour			
Environmental enrichment			
Housing:			
Free-range, shelf, cabinet, isolator			
Cage type and size			
Number and method of distribution of animals per cage			

A simple but effective Master Plan



A Contingency Plan, based upon risk assessment

- Access to emergency services (police, fire, medical and veterinary help, security guards, personnel transport in cases of acute illness)
- Means of communication with staff members at all levels
- SOPs for acute illness, including
 - serious haemorrhages
 - fainting
 - allergic and anaphylactic reactions
 - burns
 - head injuries
 - bites
 - corrosive injuries
 - and forms for reporting such injuries
- Firefighting, evacuation of personnel and animals
- Access to specialist services (e.g. ventilation system, plumbing, electrical installations, suppliers of equipment)
- Routines in cases of power failure, water leaks and (if applicable) natural disasters such as flooding
- Routines for emergency killing of animals
- Routines in cases of threats to the facility or personnel



Temporary staff at weekends and holidays

<https://norecopa.no/prepare/6-facility-evaluation/master-plan-and-sops/contingency-plan>

Contingency and redundancy

Anything that can go wrong, will go wrong (Murphy's Law)



Photo: NMBU

Consult the animal carers and technicians from Day 1:

- they have a right to know and will be more motivated
- they know the possibilities (and limitations) in the animal facility
- they often possess a large range of practical skills and are good at lateral thinking
- they know the animals best
- the animals know them best
- lack of involvement creates anxiety, depression and opposition to animal research, as well as limiting creativity which might improve the experiments

An International Culture of Care Network

norecopa.no/CoC

Before you speak...

THINK!

T - is it true?
 h - is it helpful?
 i - is it inspiring?
 n - is it necessary?
 k - is it kind?

FREE YOUR MIND and THINK

The International Culture of Care Network

Thomas Bertelsen, Novo Nordisk A/S, Denmark; **Nikolaos Kostomitsopoulos**, Biomedical Research Foundation Academy of Athens, Greece; **Anja Petrie**, University of Aberdeen, UK; **Adrian Smith**, Norecopa, Norway

Background

Recital 31 of the Directive 2010/63/EU states that breeders, suppliers and users of research animals should have an animal-welfare body which fosters a **climate of care** and provides tools for implementation of the 3Rs. Many user establishments use the phrase 'Culture of Care' on their websites, but no clear definition of this exists.

The aims of the Culture of Care Network

To provide a forum for the quick and efficient dissemination of ideas and efforts to create a culture of care

To promote a mindset and behaviour that continuously and proactively works to advance laboratory animal welfare and the 3Rs

To aim for more than a culture of compliance

To encourage a culture of challenge, rather than accepting established practice

The experience gained by the network will be useful for the review of Directive 2010/63/EU, which is due by November 2017.



Our members

The network consists of people with a large range of backgrounds:

- Laboratory animal scientists & technicians
- Laboratory animal veterinarians
- Members of Animal Welfare Bodies & National Committees
- Representatives of National competent authorities
- Communications experts
- Members of animal welfare organisations

This diversity of competency and perspectives ensures that the network encourages a culture of care both for the animals used in research and those working with them.

We are currently 28 members in 14 countries.



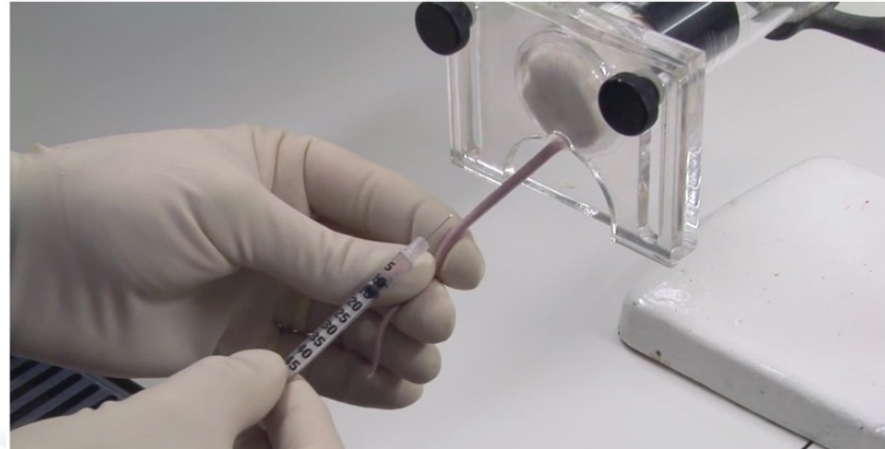
Interested in joining?

Members are expected to work actively with Culture of Care. Please contact Thomas Bertelsen (tb@novonordisk.com)

References:

- M H Lloyd, B W Foden, S E Wolfensohn. Refinement: promoting the three Rs in practice. *Laboratory Animals* 2008; 42:284-293
- J Klein, K A Bayne: Establishing a Culture of Care. *ILAR Journal* 2007; 48(1):3-11
- H Herzog: Ethical Aspects of Relationships Between Humans and Research Animals. *ILAR Journal* 2002; 43(1):27-32
- <https://norecopa.no/alternatives/culture-of-care>
- http://ec.europa.eu/environment/chemicals/lab_animals/pubs_guidance_en.htm

An example: i.v. injection of a radioactive isotope:



procedureswithcare.org.uk/intravenous-injection-in-the-mouse

norecopa.no/PREPARE

PREPARE Checklist | 1-Literature searches | 2-Legal issues |
3-Ethical issues, Harm-Benefit Assessment and humane endpoints | 4-Experimental design and statistical analysis |
5-Objectives and timescale, funding and division of labour | 6-Facility evaluation | 7-Education and training |
8-Health risks, waste disposal and decontamination | 9-Test substances and procedures | 10-Experimental animals |
11-Quarantine and health monitoring | 12-Housing and husbandry | 13-Experimental procedures |
14-Humane killing, release, re-use or re-homing | 15-Necropsy | Comparison with ARRIVE

Thanks to Norecopa's main sponsors:

- Standing Committee on Business Affairs, Norwegian Parliament
- Norwegian Ministries of Agriculture and Fisheries
- Research Council of Norway
- Laboratory Animals Ltd.
- Nordic Society Against Painful Experiments (NSMSD)
- Norwegian Society for Animal Protection
- Novo Nordisk
- Scottish Accreditation Board
- Stiansen Foundation
- Universities Federation for Animal Welfare (UFAW)
- US Department of Agriculture



SCOTTISH ACCREDITATION BOARD



Dyrebeskyttelsen Norge

FELASA, 10-13 June 2019

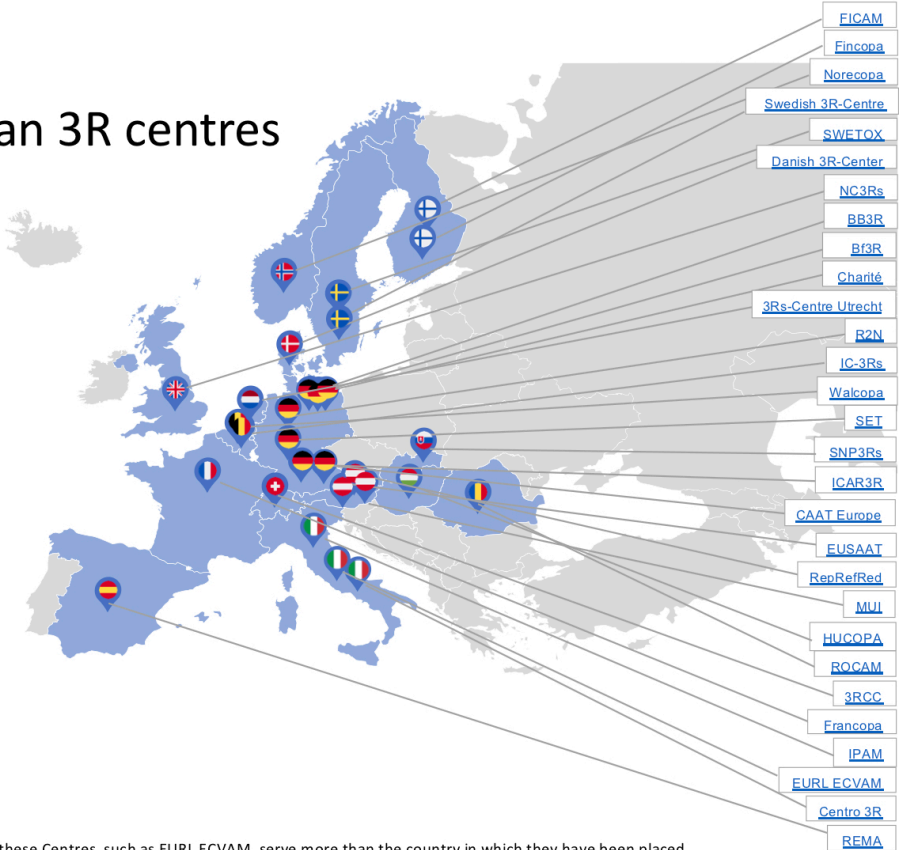
Norecopa: PREPARE for better research

Interactive map of European 3R Centres

European 3R centres

Map:
norecopa.no/3REuropeOverview

List of centres:
norecopa.no/3REurope



Please note that some of these Centres, such as EURL ECVAM, serve more than the country in which they have been placed.
 This overview has been compiled by Norecopa. Please report any errors or send suggestions for additions to post@norecopa.no
 Designed by PresentationGo.com. Flags from flaticon.com

Mild, Moderate or Severe? A compilation of severity classification



norecopa.no/severity

COMPILATION OF SEVERITY CLASSIFICATIONS ACROSS EUROPE

I Interventions on body systems and functions	II Induction of diseases	III Pharmacology, other external causes	IV Housing, environment and behaviour	V Fetuses and premature animals	VI Clinical signs
Substance administration Specimen collection 1. Collection of body fluids 2. Tissue sampling Surgical interventions 1. Anaesthesia 2. Surgical interventions in general 3. Abdominal and chest cavity 4. Musculoskeletal system 5. Implantation of mini-pumps, transponders 6. Organ transplantation 7. Implanted probes 8. Others	Heart & circulation 1. Heart 2. Circulation 3. ECG measurements Infectious diseases 1. Infections in general 2. Zoonosis 3. Bacterial infections 4. Viral infections 5. Mycotic infections Neurology & sensory organs 1. Convulsions 2. CNS lesions 3. Ischemias 4. Visual system Endocrine, nutritional & metabolic diseases 1. Endocrinology 2. Bone metabolism 3. Glucose metabolism 4. Body weight loss Neoplasms Immunology 1. Transplantation 2. Cellular reactions 3. Autoimmune reactions 4. Immunisation 5. Immunity 6. Inflammation 7. Arthritis 8. Asthma	Physical impacts 1. Irradiation & chemotherapy 2. Exposure to electricity 3. Exposure to heat 4. Exposure to cold 5. Exposure to acids & bases 6. Traumatization 7. Pressure change 8. Sound & ultrasound 9. Magnetic fields 10. Chronic hypoxia Generation of pain Pharmacological studies 1. Toxicity studies - general 2. Acute toxicity 3. Sub-acute toxicity 4. Chronic toxicity 5. Reproduction toxicology 6. Pharmacokinetic studies 7. Batch testing	Housing & Nutrition 1. Housing in general 2. Gnotobiology 3. Use of metabolic cages 4. Deprivation - water 5. Deprivation - food 6. Exposure to overstimulation Breeding & Reproduction 1. Identification method and tissue sampling for genotyping 2. Germ cells GA animals 1. GA animal model in the experiment 2. Phenotype characteristics 3. Generation of GA animals 4. Tissue sampling for genotyping Behaviour 1. Aversive learning, conditioned avoidance behaviour and conflict tests 2. Deprivation - social 3. Deprivation - sleep 4. Deprivation - motion & mobility 5. Pharmacologically induced behavior 6. Exercise 7. Specific behavior tests 8. External telemetry devices	1. Feeding 2. Surgical interventions 3. Reproductive toxicology	1. Condition of fur, grooming, body orifices 2. Breathing, heart rate 3. Body temperature 4. Behaviour & body posture 5. Reaction to artificial stimulation, handling, behaviour, reaction to physical contact 6. Food & water consumption 7. Prostration 8. Self trauma 9. Body weight

Poster PF12

Anne Zintzsch, Nikolaos Kostomitsopoulos & Jan-Bas Prins

'One Europe': The challenge of consistency in severity classification
 Anne Zintzsch¹, Adrian Smaliv², Nikolaos Kostomitsopoulos³ & Jan-Bas Prins⁴
¹ DR Centre for Animal Welfare, University of Göttingen & Proficiency for Laboratory Animal Science and Animal Welfare, Germany; ² Norecopa, Norway; ³ Biomedical Research Foundation of the Academy of Athens, Greece; ⁴ The Francis Crick Institute, UK & London University Medical Centre, The Netherlands

<https://norecopa.no/severity>


Background
 Severity classification is an important factor in the project authorization of animal experiments and... (text continues)

Compilation as the first step
 In order to be able to compare results across different studies, we have collected existing guidelines and compiled them into a table... (text continues)

A compilation of severity classifications is available online

The way forward
 The next step is to review the severity classifications, add more guidance notes and include more examples of severity classifications... (text continues)

<https://norecopa.no/severity>




Norecopa: PREPARE for better research

FELASA, 10-13 June 2019

English-language newsletters

Contact oss
+47 41 22 09 49
post@norecopa.no

 Norecopa on Facebook

Street address
Ullevålsveien 68
0454 Oslo

Postal address
% Norwegian Veterinary
Institute
P.O. Box 750 Sentrum
N-0106 Oslo, Norway

Org.no. 992 199 199
Bank account: 7694 05 12030
(IBAN: NO51 7694 0512 030)
(payment must be marked
'12025 Norecopa')

Shortcuts


- > Give us some feedback!
- > 2010/63/EU
- > Information material
- > Norecopa's Board
- > Secretariat
- > Sponsors
- > Cookies & Privacy
- > Site map


Subscribe to our newsletter

Your email address:

> Browse our latest newsletters

Resources developed in collaboration with:

 Norges miljø- og
biovitenskapelige
universitet

 U.S. Department
of Agriculture



norecopa

"We ARRIVED, because we were PREPARED"

norecopa.no/prepareFELASA

Norecopa: PREPARE for better research

FELASA, 10-13 June 2019